

Literature-derived Bioaccumulation Models for Energetic Compounds in Plants and Soil Invertebrates

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EXECUTIVE SUMMARY

Concentrations of contaminants in tissue are needed to estimate exposure of wildlife. In the absence of measured data, models are used. Soil-to-plant tissue bioaccumulation models for 10 energetic compounds [HMX, perchlorate, RDX, tetryl, TNB, TNT, and its metabolites (2-ADNT, 4-ADNT, 2,4-DNT, and 2,6-DNT)] representing 4 plant tissue types (leaf, root, fruit, and seed), and soil-to-whole-body earthworm bioaccumulation models for 8 energetic compounds [HMX, perchlorate, RDX, TNT, and its metabolites (2,4-DANT, 2,6-DANT, 2-ADNT, and 4-ADNT)] were developed. Models were developed based on data extracted from published, peer-reviewed literature and gray literature. Data were pooled by chemical, taxonomic groups, and tissue types to determine whether linear relationships between chemical concentrations in soil and in biota (plant tissues and earthworms) existed. All together, 36 soil-to-tissue combinations were analyzed and presented in this report.

For plants, six of ten chemicals (2-ADNT, 4-ADNT, 2,6-DNT, HMX, RDX, and TNT) had significant natural log-linear relationships between soil to at least one plant tissue type. Whereas positive relationships were observed for 2-ADNT, 2,6-DNT, HMX, and RDX, inverse relationships were obtained for TNT in soil to plant roots and 4-ADNT in soil to plant foliage. Summary statistics of BAFs (ratio of soil to tissue) were generated for all chemical-tissue combinations. Only 2,4-DNT and 4-ADNT had median soil-to-plant foliage BAFs of less than one; the remaining chemicals (2,6-DNT, 2-ADNT, HMX, perchlorate, RDX, tetryl, TNB, and TNT) all had median foliage uptake values greater than one. Three chemicals had soil-to-fruit uptake values (RDX, tetryl, and TNT). Among these three analytes, RDX was the only chemical that had a median soil-to-fruit uptake value of greater than one. Median soil-to-root uptake values for RDX, tetryl, and TNT were greater than one. Among the four chemicals with uptake data for seeds (perchlorate, RDX, tetryl, and TNT), only RDX and tetryl had median uptake values greater than one.

For soil-to-earthworm, four chemicals (RDX, perchlorate, 2-ADNT, and 4-ADNT) had statistically significant log-linear relationships between soil to earthworms. Whereas RDX, 2-ADNT, and 4-ADNT displayed positive relationships, that for perchlorate was negative. Summary statistics of BAFs (ratio of soil to earthworms) were generated for all analytes. Five chemical pairs: HMX in soil to HMX in earthworms, TNT-to-TNT, perchlorate-to-perchlorate, TNT-to-2ADNT, and TNT-to-4ADNT had median soil-to-tissue BAFs of less than one; the remaining chemical pairs: HMX/HMX, perchlorate/perchlorate, RDX/RDX, TNT/2-ADNT, TNT/2,4-DANT, 2,4-DANT/2,4-DANT, 2,6-DANT/2,6-DANT, 2ADNT/2ADNT, and 4ADNT/4ADNT all had median uptake values of greater than one.

1. INTRODUCTION

Estimation of the risks that soil contamination presents to herbivorous and vermivorous (e.g., earthworm-eating) wildlife requires measuring the contaminant concentration in terrestrial biota. These data may be acquired either by direct measurement or estimation. Direct measurement consists of collecting and analyzing contaminant concentrations in biota tissues from contaminated sites. Because this approach provides information on the actual contaminant loading in on-site earthworms, direct measurement contributes the least uncertainty to exposure estimates and is therefore the preferred approach. However, for various reasons (incompatible sampling schedule; insufficient time, personnel, or finances to support field sampling, etc.), direct measurement may not be feasible. When direct measurement of contaminants is not possible, estimation is the only alternative.

Contaminant loads in terrestrial biota may be estimated using uptake factors (UFs) or empirically derived bioaccumulation models. UFs, the ratios of contaminant concentrations in biota to those in soil, are the simplest method for estimating contaminant loads in biota. In practice, if the contaminant concentration in soil is known (likely in almost all retrospective ecological risk assessments), the concentration in biota may be estimated by multiplying the soil concentration by the UF. The use of UFs depends on the assumption that the concentration of chemicals in organisms is a linear, no-threshold function of concentrations in soil. This is expected to be the case for xenobiotic chemicals like PCBs that are passively accumulated and not metabolized to any significant extent. It will not be the case if the chemical in question is well-regulated by the organism, either because it is an essential nutrient or because it is a toxicant with effective inducible mechanisms for metabolism or excretion. Such regulated chemicals will, within the effective concentration range for the mechanism, have nearly constant concentrations in biota regardless of soil concentrations, except at deficient concentrations. Various complex patterns are also possible due to lack of induction at low concentrations, saturation kinetics at high concentrations, toxicity at high concentrations, or other processes. Despite these situations that lead to violation of the assumptions, UFs are commonly used in risk assessments.

Regression models are another approach to estimating contaminant concentrations in biota. These models are generally simple linear or log-linear regressions of the soil contaminant concentration on the biota concentration. While there has been considerable research concerning the uptake of soil contaminants by plants and earthworms, most studies use data from a limited number of locations and focus on a limited number of analytes. The purpose of this report was to assemble two sets of database of soil-to-earthworm and soil-to-plant contaminant concentration data from published literature, develop UFs and linear regression bioaccumulation models from these data for energetic compounds.

In this report, both UFs and regression models were developed, because, while regression models are most likely to consistently provide the best estimate of biota body burdens, UFs are required by some regulatory agencies. In addition, when no regression model fits the uptake data well, a conservative UF may be employed in screening assessments to

determine whether site-specific studies are needed. The models presented in this report will facilitate the more accurate estimation of contaminant exposure experienced by herbivores and earthworm-consuming wildlife.

2. DATABASE DEVELOPMENT

2.1. Plant Bioaccumulation Database

This investigation evaluated data for 10 energetic compounds with soil-to-plant tissue uptake studies representing 4 plant tissues (leaf, root, fruit, and seed) and resulted in 25 soil-to-tissue combinations. The 10 energetic compounds were: HMX, perchlorate, RDX, Tetryl, TNB, TNT and its metabolites (2-ADNT, 4-ADNT, 2,4-DNT, and 2,6-DNT) (Table 2.1-1). Studies were primarily from greenhouse pot studies in which chemicals were added to soil.

2.1. Earthworm Bioaccumulation Database

The soil invertebrate bioaccumulation dataset consisted entirely of earthworm species from 3 different genera: Aporectodea, Eisenia, and Lumbricus. Our investigation covered 8 energetic compounds [HMX, perchlorate, RDX, TNT, and its metabolite (2,4-DANT, 2,6-DANT, 2-ADNT, and 4-ADNT)] in whole-body tissues of earthworms. Because some of the studies measured TNT metabolites in earthworms, the database contained a total of 11 soil-to-earthworm combinations that arised from parent compounds in soil to parent or metabolites in earthworm. Studies were primarily from laboratory investigations in which chemicals were added to soil.

2.2 Data Treatment

Soil and tissue concentrations were expressed in dry-weight basis (and converted to dry weight if the reported concentration were originally in wet weight concentrations). As a result, summary statistics and regression analysis were all based on dry weight basis for soil and biota concentrations. Perchlorate studies were frequently performed on perchlorate salts and expressed as salt concentrations. To facilitate comparisons os data from different studies, concentrations were all converted to perchlorate anions. Although most studies reported that plant material was washed, studies were not excluded if the extent of washing was not stated in the paper. Studies were used even if the individual investigators observed no correlation between concentrations of contaminants in soils and plants. Concentrations of chemicals in soil or plants were sometimes estimated visually from a figure if contacts with authors were unsuccessful. Measured concentrations were preferred over nominal soil concentrations, but nominal concentrations were included in the database if measured concentration were not available. Soil and biota measurements were quantified by HPLC coupled with UV detector or GC/MS; only one study quantified chemical concentrations using HPLC alone. Thus, quantitation measurements were considered adequate.

2.3 Organization of the Database: Plant and Earthworm Dataset

The bioaccumulation database which includes soil and biota (plant and earthworm) concentrations, soil parameters, exposure time, chemical form, dry or wet weight, extraction method, and species information are presented in Appendix A-1 (plant) and A-2 (earthworm). Each plant and earthworm species, taxonomic classification, soil type, location, concentration of the test chemical in soil, and form of an added chemical represents an independent observation in the dataset. For plants, differences in above-ground plant tissue (e.g. seed vs. leaf) were considered as separate soil-to-plant tissue combinations. Concentrations of contaminants in soil at the time of biota sampling were used if known because it most closely represents soil concentrations and plant tissue concentrations at a site and for a site-specific risk assessment. If these final concentrations were not measured (as was often the case in greenhouse pot studies), the initial concentration of the chemical measured in soil or nominal soil concentration was assumed to be equivalent to the final concentration. It was assumed that the soils used were free from contaminants prior to the addition of test compounds. In field experiments where soil and tissues of resident biota were analyzed, the change in soil concentration of a chemical over time was assumed to have reached equilibrium.

Detailed summaries of experimental conditions, analytical methods used, and relevant notes for the purpose of this investigation were summarized for each data source and presented in Appendix B-1 (for plants) and B-2 (for earthworms).

2.4 Data Analysis: Plant and Earthworm BAFs

To evaluate if a linear relationship between the chemical concentration in soil and that in terrestrial biota existed, simple linear regressions were performed using SAS PROC REG (SAS Inst. Inc. 1999). Chemical concentrations in both soil and biota tissues were transformed to natural-log (ln) prior to regression analyses. Because data concerning the number of individuals included in composites or means were not available for all observations, no weighting of observations was applied. Summary statistics of BAF values (min, max, median, and 90th percentile) were provided. For plants, a distinction was made in the dataset between monocot and dicot species, and analysis of dicot and monocot data were analyzed separately as well as pooled in the summary and ln-ln regression analysis.

3. RESULTS OF BIOACCUMULATION MODELS

3.1. Soil-to-Plant Tissue BAF Models

Table 3.1-1 presented a summary of soil-to-plant tissue BAF values and the result of the log-log regression model. The level of significance was based on alpha of 0.05.

The results of the bioaccumulation analyses are summarized from Table 3.1-1 based on pooled monocot and dicot (indicated as "all" in the database) as follows:

- Among all the chemicals examined, only two chemicals (2,4-DNT and 4-ADNT) have a median soil-to-plant foliage uptake of less than one; the remaining chemicals (2,6-DNT, 2ADNT, HMX, perchlorate anion, RDX, Tetryl, TNB, and TNT) all have median uptake values of greater than one.
- Only 3 chemicals have soil-to-fruit uptake values (RDX, tetryl, and TNT). RDX is the only chemical that have a median uptake value of soil-to-fruit greater than one.
- Six chemicals have soil-to-root uptake values (2-ADNT, 4-ADNT, HMX, RDX, Tetryl, and TNT). Tetryl and TNT have median uptake values greater than one.
- Of the four chemicals with uptake data from soil to seed (perchlorate, RDX, Tetryl, and TNT), only RDX and Tetryl have median uptake values greater than one.
- RDX has median uptake values for all plant tissue compartments (foliage, fruit, and seed), except for root at a median value of 0.73.

Six chemicals (2-ADNT, 2,6-DNT, HMX, RDX, and TNT) have significant log-linear relationships between soil to at least one plant tissue compartment (Table 3.1-1). All were positive with the exception of TNT and 4-ADNT, which displayed inversely proportional relationships. Figure 3.1-1 through 3.1-25 presented the scatter plots and regression models (when applicable) of the soil-to-plant tissue data. Results of the log-log regression analyses for soil-to-plant bioaccumulation are summarized as follows:

- Five chemicals (2-ADNT, 4-ADNT, HMX, RDX, and perchlorate) have significant log-linear relationship between soil to at least one plant tissue compartment when plant species are separately analyzed by monocots and dicots.
- Six chemicals (2-ADNT, 4-ADNT, HMX, perchlorate, RDX, and TNT) have significant log-linear relationships between soil to at least one plant tissue compartment when monocot and dicot plant uptake data are pooled.
- For chemicals with statistically significant relationships, concentrations in tissue are proportional to chemicals in soil concentration. However, linear relationship between plant tissue concentration and soil concentrations are inversely proportional for TNT in soil to TNT in plant root, and for 4-ADNT in soil to 4-ADNT in foliage.

3.1. Soil-to-Earthworm BAF Model

Table 3.1-2 presents a summary of soil-to-earthworm BAF values and the results of the log-log regression analyses. The level of significance was based on alpha of 0.05.

Among all chemicals examined, 5 chemicals pairs: HMX in soil to HMX in earthworms, perchlorate-to-perchlorate, TNT-to-TNT, TNT-to-2ADNT, and TNT-to-4ADNT had median soil-to-tissue BAFs of less than one. The remaining 6 chemical pairs (2ADNT/2ADNT, 4ADNT/4ADNT, RDX/RDX, TNT/2,4-DANT, 2,4-DANT/2,4-DANT, and 2,6-DANT/2,6-DANT] all had median uptake values of greater than one.

Four chemicals (RDX, perchlorate, 2-ADNT, and 4-ADNT) had statistically significant log-linear relationships between soil and earthworms; inversely proportional relationships were found for perchlorate. Figures 3.2-1 through 3.2-11 present the scatter plots and regression models (when applicable) of the soil-to-earthworm data.

4. REFERENCE

Cataldo, D.A., S.D. Harvey, R.J. Fellows. 1993. The Environmental Behavior and Chemical Fate of Energetic Compounds (TNT, RDX, Tetryl) in Soil and Plant Systems. Presented at the 17th Annual Army Environmental R&D Symposium and 3rd USACE Innovative Technology Transfer Workshop. June 22-24. Williamsburg, VA.

Folsom, B. L. Jr., J.C. Pennington, C.L. Teeter, M.R. Barton, J.A. Bright. 1988. Effects of Soil pH and Treatment Level on Persistence and Plant Uptake of 2,4,6-Trinitrotoluene. Technical Report EL-88-22. US Army Engineer Waterways Experiment Station. Vicksburg, MS.

Groom, C.A, A. Halasz, L. Paquet, N. Morris, L. Olivier, C. Dubois, and J. Hawari. 2002. Accumulation of HMX (Octahydro-1,3,5,7-tetranitro-1,3,5,7-tetrazocine) in indigenous and agricultural plants grown in HMX-contaminated anti-tank firing-range soil. Environ. Sci. Technol. 36:112-118.

Johnson, M.S., L.S. Franke, R.B. Lee, and S.D. Holladay. 1999. Bioaccumulation of 2,4,6-trinitrotoluene and polychlorinated biphenyls through two routes of exposure in a terrestrial amphibian: Is the dermal route significant? 18:873-878.

Lachance, B., S. Rocheleau, J. Hawari, P. Gong, F. Leduc, J. Apte, M. Sarrazin, M. Martel, G. Bardai, S. Dodard, and G. I. Sunahara. 2003. Bioaccumulation of Nitro-Heterocyclic and Nitroaromatic Energetic Materials in Terrestrial Receptors in a Natural Sandy Loam Soil. Biotechnology Research Institute, National Research Council Canada. Montreal, Quebec. Canada.

Lachance, B., A.Y. Renoux, M.Sarrazin, J. Hawari, G.I. Sunahara. 2004. Toxicity and bioaccumulation of reduced TNT metabolites in the earthworm *Eisenia andrei* exposed to amended forest soil. Chemosphere. 55:1339-1348.

Parsons Engineering. 2001. Scientific and Technical Report for Perchlorate Biotransport Investigation (Interim Final): A Study of Perchlorate Occurrence in Selected Ecosystems. Austin, TX.

Price, R.A., J.C. Pennington, S.L. Larson, D. Neumann, C. A. Hayes. 1997. Plant Uptake of Explosives from Contaminated Soil and Irrigation Water at the Former Nebraska Ordnance Plant, Mead, Nebraska. Technical Report EL-97-11. US Army Corps of Engineers, Waterway Experiment Station, Vicksburg, MS.

Renoux, A.Y., M. Sarrazin, J. Hawari, G.I. Sunahara. 2000. Transformation of 2,4,6-trinitrotoluene in soil in the presence of the earthworm *Eisenia andrei*. Environ. Toxicol. Chem. 19:1473-1480.

Robidoux, P.Y., C. Svendsen, M. Sarrazin, S. Thiboutot, G. Ampleman, J. Hawari, J.M, Weeks, G.I. Sunahara. 2004. Assessment of a 2,4,6-trinitrotoluene-contaminated site using *Aporrectodea rosea* and *Eisenia andrei* in Mesocosms. Arch. Environ. Contam. Toxicol. 28:56-67.

SAS Inst. Inc. 1999. SAS/STAT User's Guide, Version 8. SAS Institute, Cary, NC.

Schneider, J.F., S.D. Zellmer, N.A. Tomczyk, J.R. Rastorfer, D. Chen, and W.L. Banwart. 1995. Uptake of Explosives from Contaminated Soil by Existing Vegetation at the Iowa Army Ammunition Plant. ADA296529. Prepared by Argonne National Laboratory, Argonne, IL for US Army Environmental Center. Aberdeen Proving Ground, MD.

Zellmer, S.D. J.F. Schneider, N.A. Tomczyk, W.L. Banwart, and D. Chen. 1995. Plant Uptake of Explosives from Contaminated Soil at the Joliet Army Ammunition Plant. ADA 296529. US Army Environmental Center, Aberdeen Proving Ground, MD.

Table 2.1-1. Full Name and Short Name of Energetic Compound

CAS NO.	Full Name	Short Name
121-14-2	2,4-dinitrotoluene	2,4-DNT
606-20-2	2,6-dinitrotoluene	2,6-DNT
35572-78-2	2-amino-4,6-dinitrotoluene	2-ADNT (2-amino-DNT)
19406-51-0	4-amino-2,6-dinitrotoluene	4-ADNT (4-amino-DNT)
2691-41-0	Octahydro-1,3,5,7-tetranitro-1,3,5,7-tetrazocine	HMX
14797-73-0	Perchlorate	Perchlorate
121-82-4	Hexahydro-1,3,5-trinitro-1,3,5-triazine	RDX
479-45-8	2,4,6-trinitrophenylmethylnitramine	Tetryl
99-35-4	1,3,5-trinitrobenzene	TNB
118-96-7	2,4,6-trinitrotoluene	TNT

Table 3.2-1. Summary Statistics of Uptake Values and Regression Models for Soil Invertebrates

Figure No.	Parent Analyte	Compound in Plant Tissue	Tissue	BAF Summary Statistics						Range of Detected Concentrations				Root Mean Square Error	p	r2
				n	mean	std	min	median	max	Soil	Plant Tissue	Intercept	Slope			
Figure 3.2-1	2,4-DANT	2,4-DANT	Whole-body	2	3.71	1.98	2.31	3.71	5.10	2.17 - 32.4	5.02 - 165	0.608	1.294	0	.	.
Figure 3.2-2	2,6-DANT	2,6-DANT	Whole-body	1	3.16		3.16	3.16	3.16	44.5	140	4.945	0.000	0	.	.
Figure 3.2-3	2ADNT	2ADNT	Whole-body	21	5.37	6.01	0.0117	4.33	29.6	0.387 - 109	0.14 - 591	1.593	0.766	1.78	0.0125	0.286
Figure 3.2-4	4ADNT	4ADNT	Whole-body	22	4.15	2.61	0.0125	3.78	9.13	0.731 - 91	0.15 - 414	1.701	0.696	1.73	0.0473	0.1826
Figure 3.2-5	HMX	HMX	Whole-body	3	0.469	0.473	0.0925	0.313	1.00	9 - 400	9 - 37	1.427	0.380	0.189	0.1165	0.9337
Figure 3.2-6	Perchlorate	Perchlorate	Whole-body	4	2.26	4.13	0.0196	0.281	8.46	3.82 - 591	11.4 - 32.3	3.474	-0.198	0.363	0.0148	0.655
Figure 3.2-7	RDX	RDX	Whole-body	4	4.32	4.79	0.675	2.63	11.4	11 - 400	125 - 283	4.376	0.230	0.202	0.0067	0.812
Figure 3.2-8	TNT	2,4-DANT	Whole-body	2	1.99	2.39	0.307	1.99	3.68	1.36 - 2.73	0.836 - 5.02	2.413	-2.585	0	.	.
Figure 3.2-9	TNT	2ADNT	Whole-body	9	3.74	6.09	0.320	0.493	14.5	1.36 - 135	1.44 - 157	1.562	0.521	1.35	0.1037	0.333
Figure 3.2-10	TNT	4ADNT	Whole-body	9	10.1	19	0.560	0.678	50.6	1.36 - 135	2.52 - 113	2.718	0.259	1.39	0.3948	0.1051
Figure 3.2-11	TNT	TNT	Whole-body	6	0.0649	0.0661	0.000333	0.0581	0.170	60 - 367	0.02 - 19.7	-9.591	2.119	2.71	0.3416	0.2252

Notes:

Rows that are highlighted indicate log-log linear relationship are statistically significant ($\alpha=0.05$)

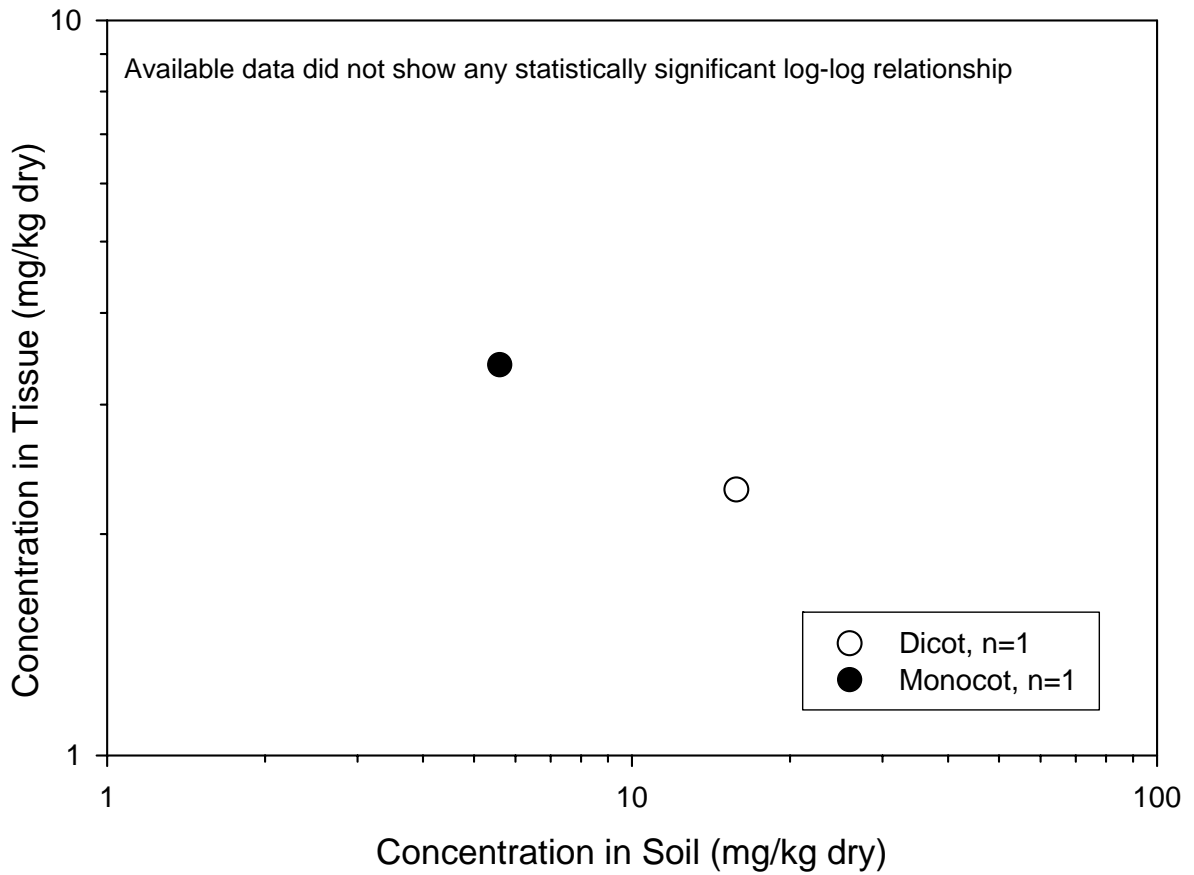


Figure 3.1-1. 2,4-DNT in Soil to 2,4-DNT in Plant Foliage

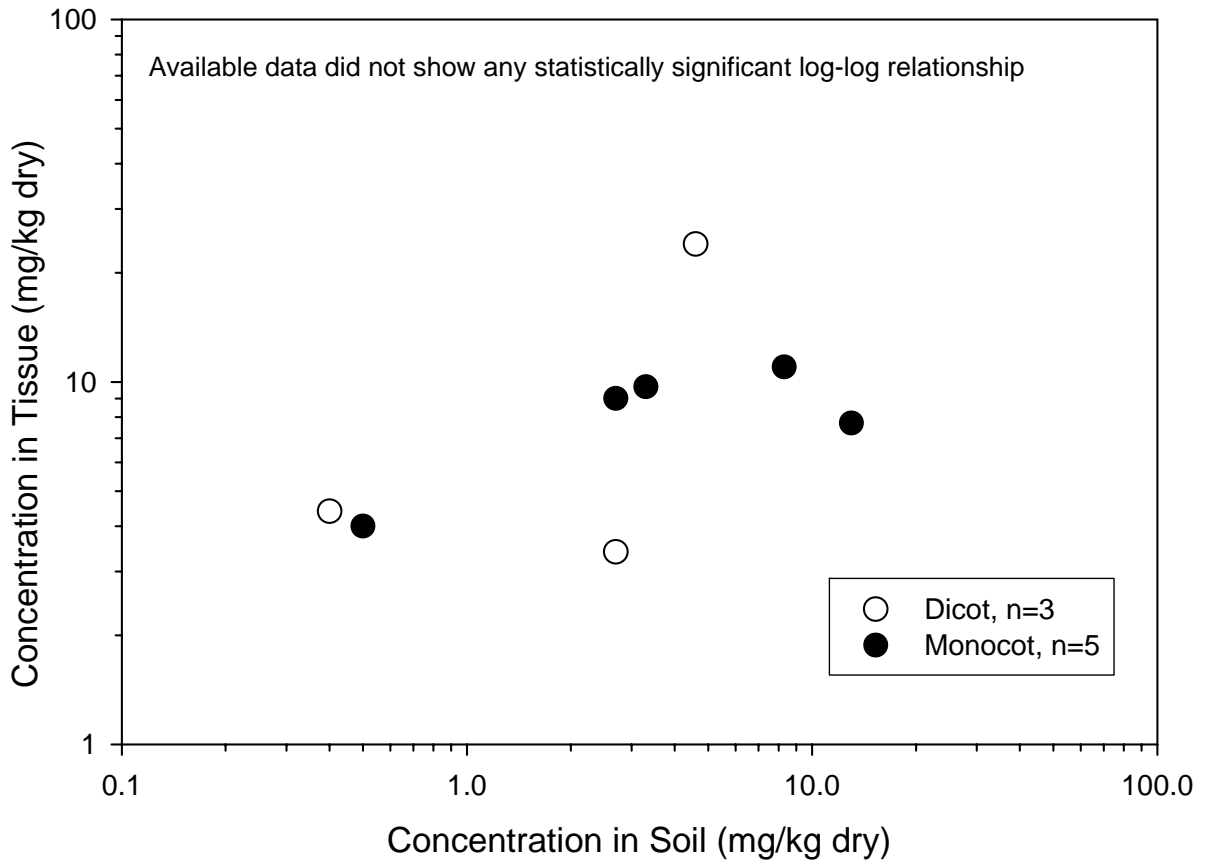


Figure 3.1-2. 2,6-DNT in Soil to 2,6-DNT in Plant Foliage

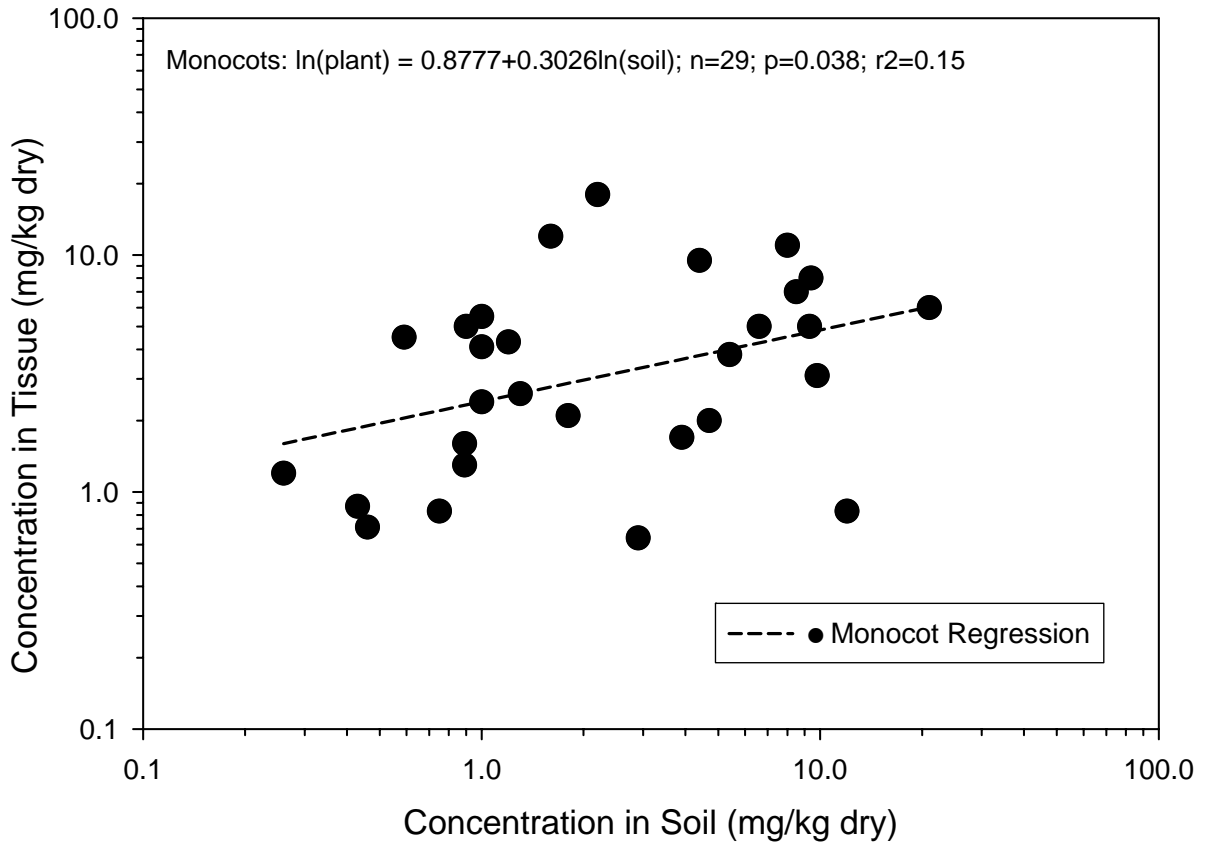


Figure 3.1-3. 2-ADNT in Soil to 2-ADNT in Plant Foliage

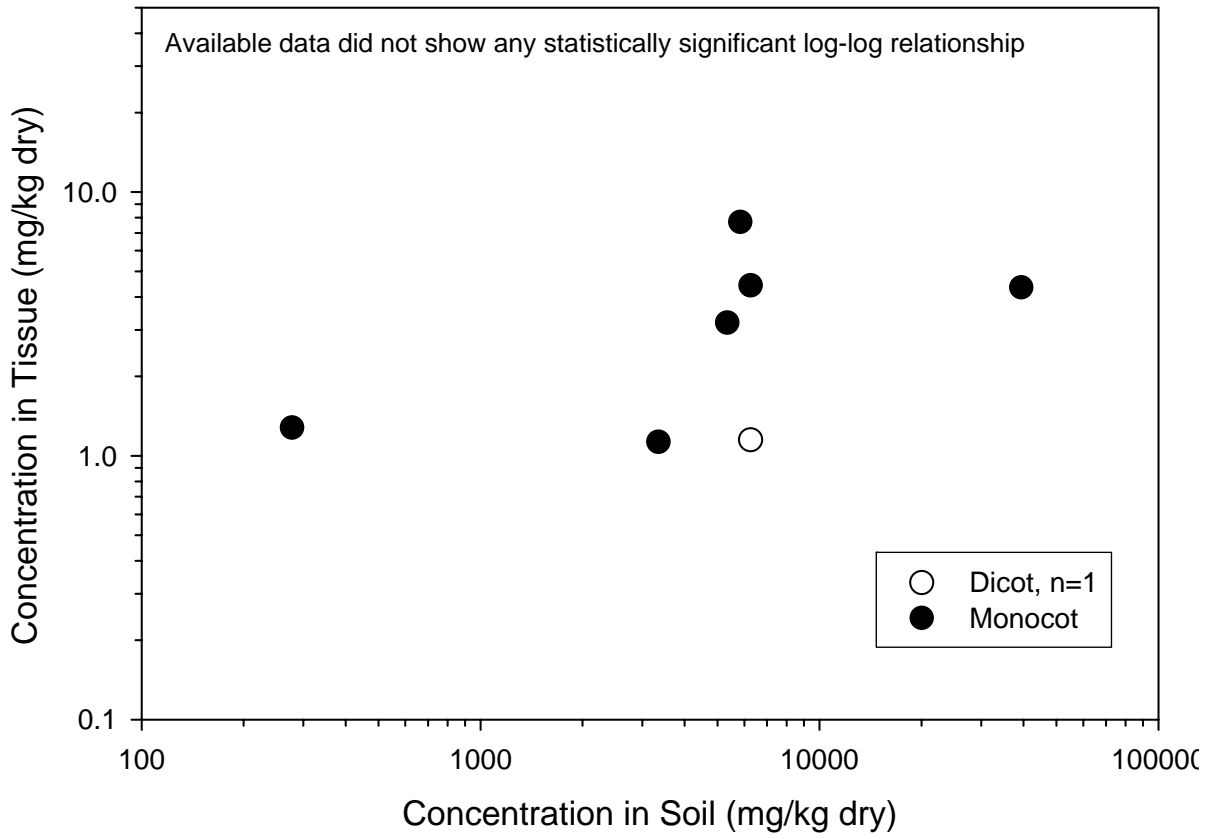


Figure 3.1-4. 2-ADNT in Soil to 2-ADNT in Plant Root

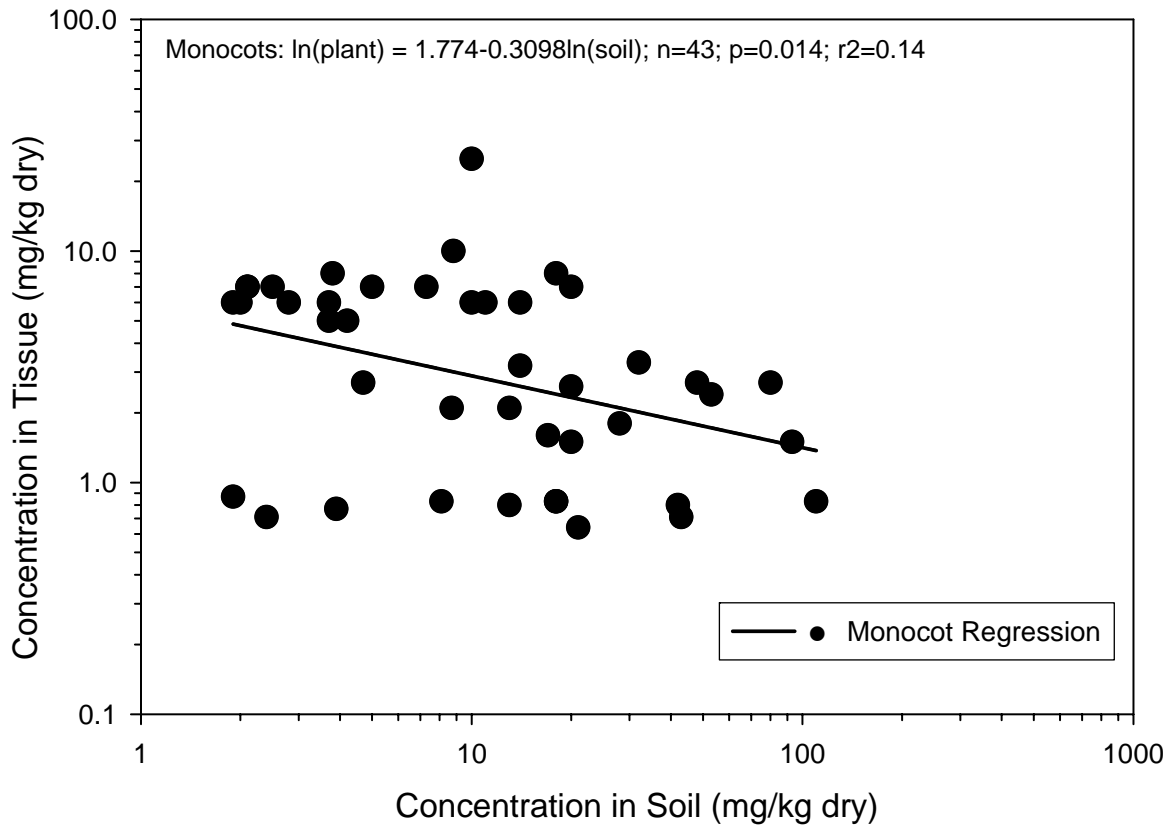


Figure 3.1-5. 4-ADNT in Soil to 4-ADNT in Plant Foliage

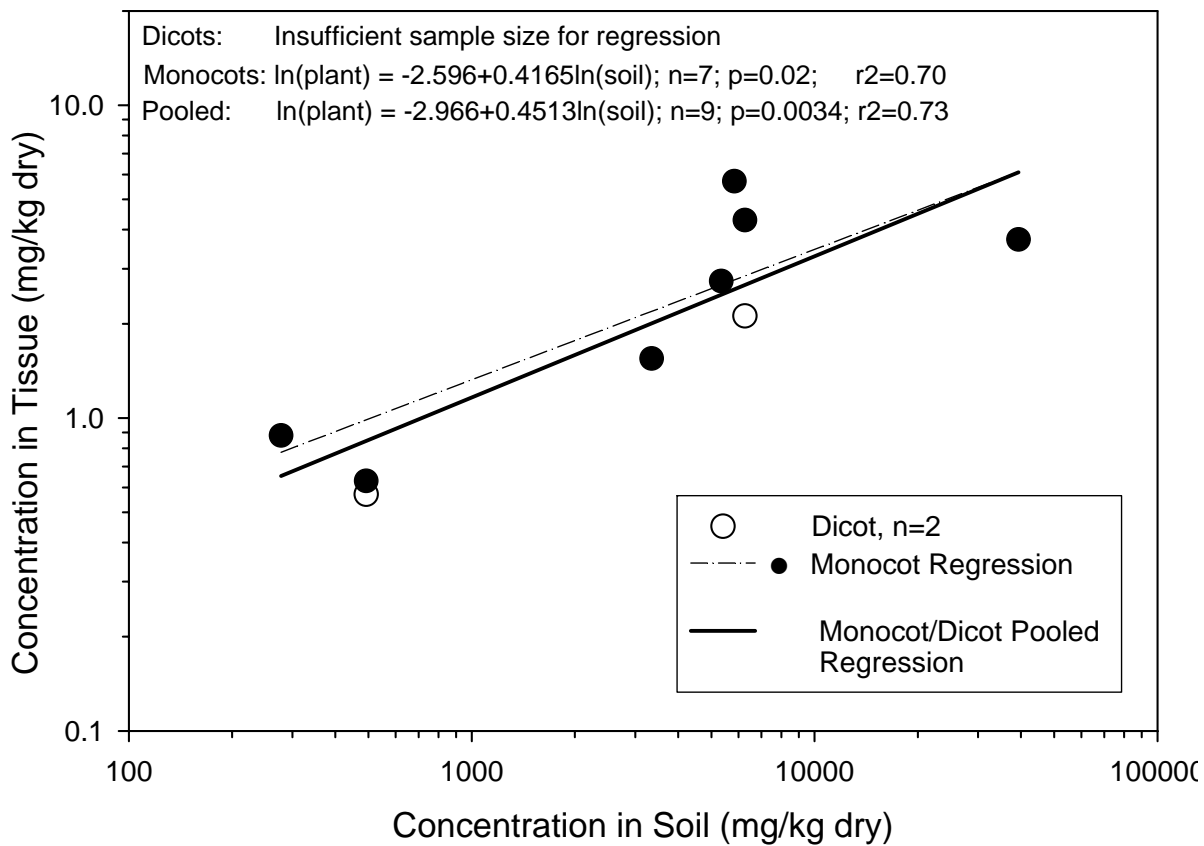


Figure 3.1-6. 4-ADNT in Soil to 4-ADNT in Plant Root

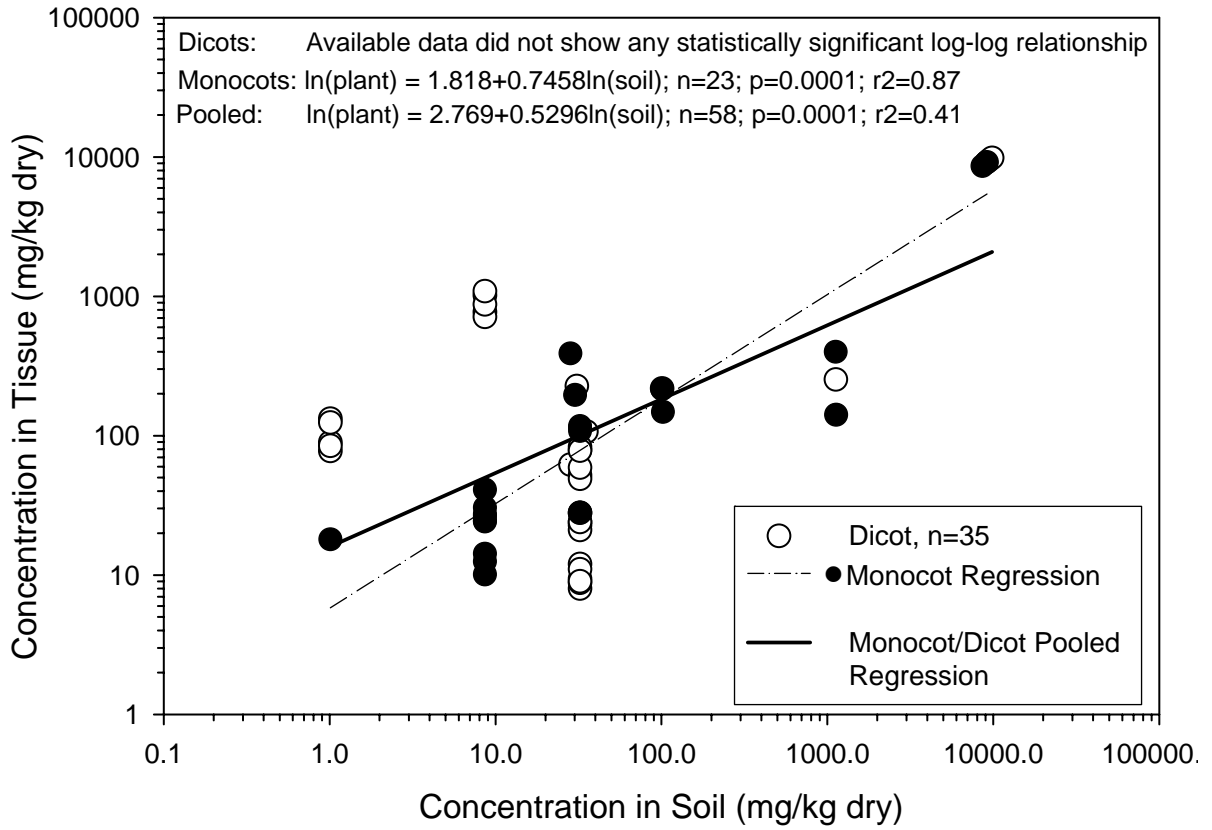


Figure 3.1-7. HMX in Soil to HMX in Plant Foliage

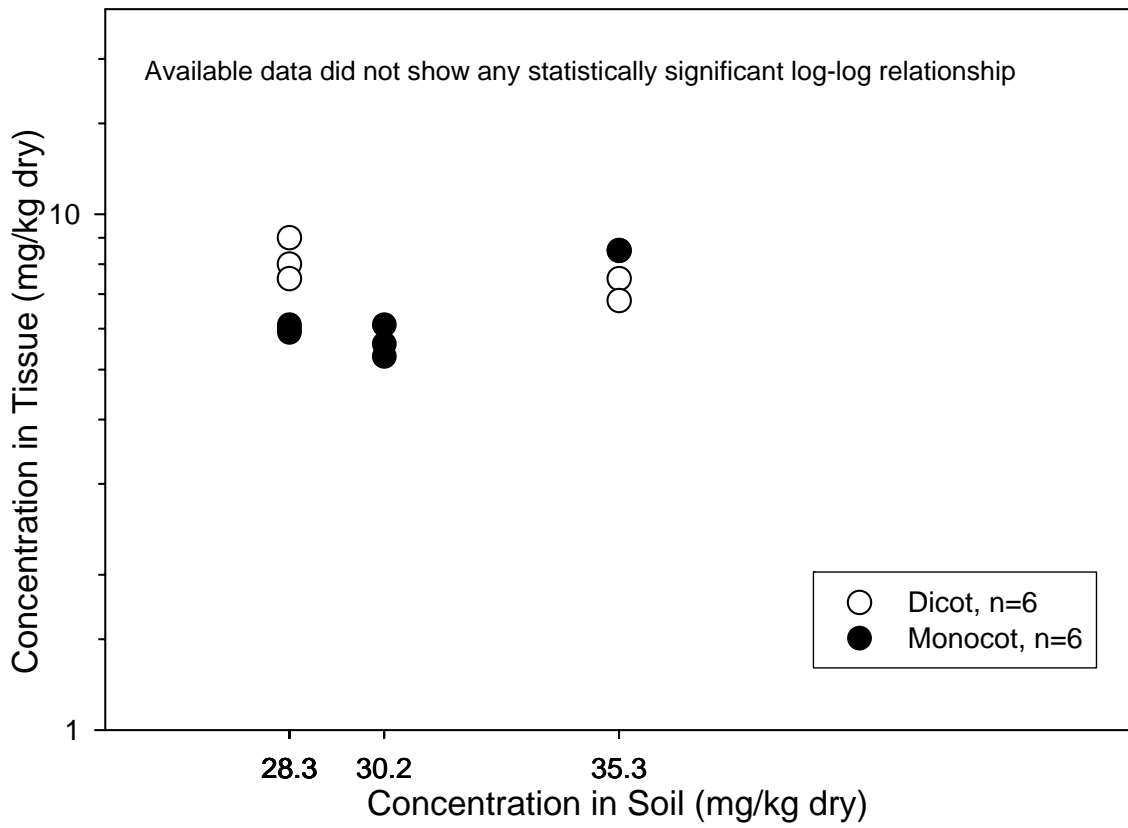


Figure 3.1-8. HMX in Soil to HMX in Plant Root

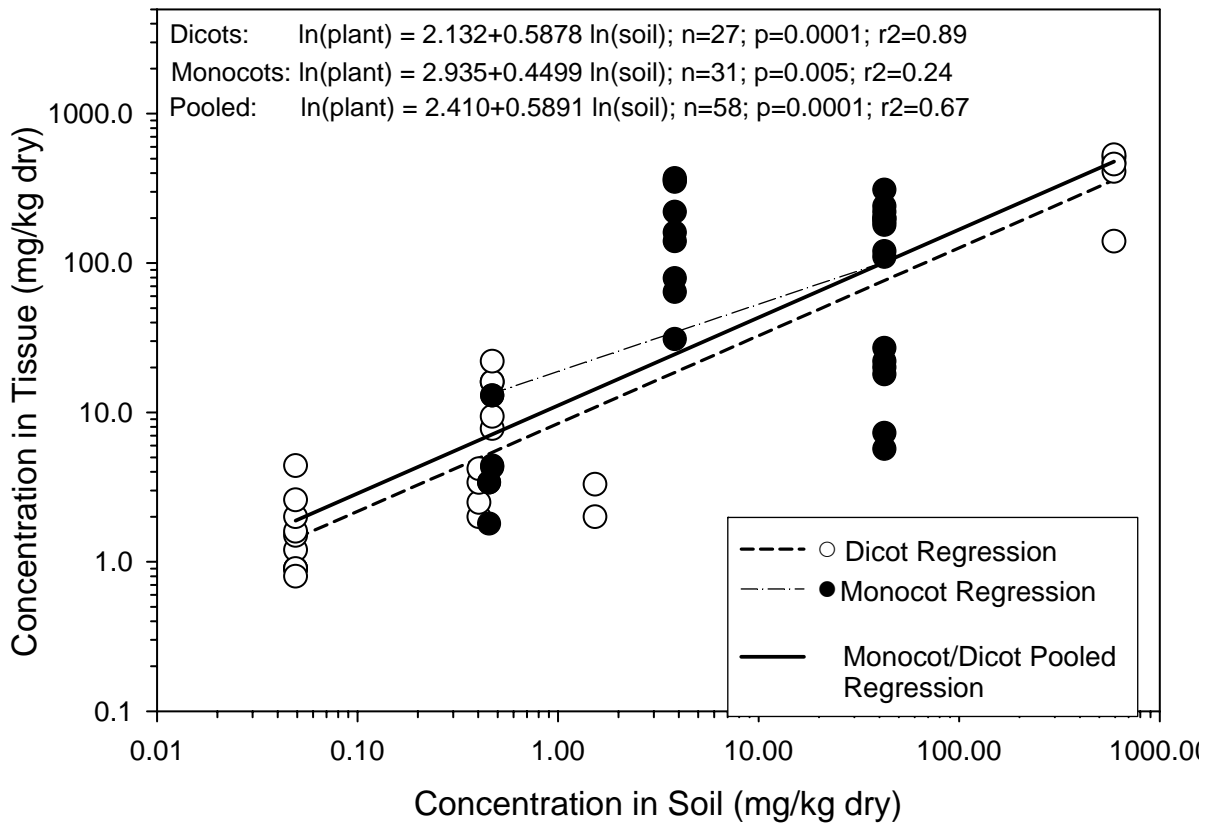


Figure 3.1-9. Perchlorate in Soil to Perchlorate in Plant Foliage

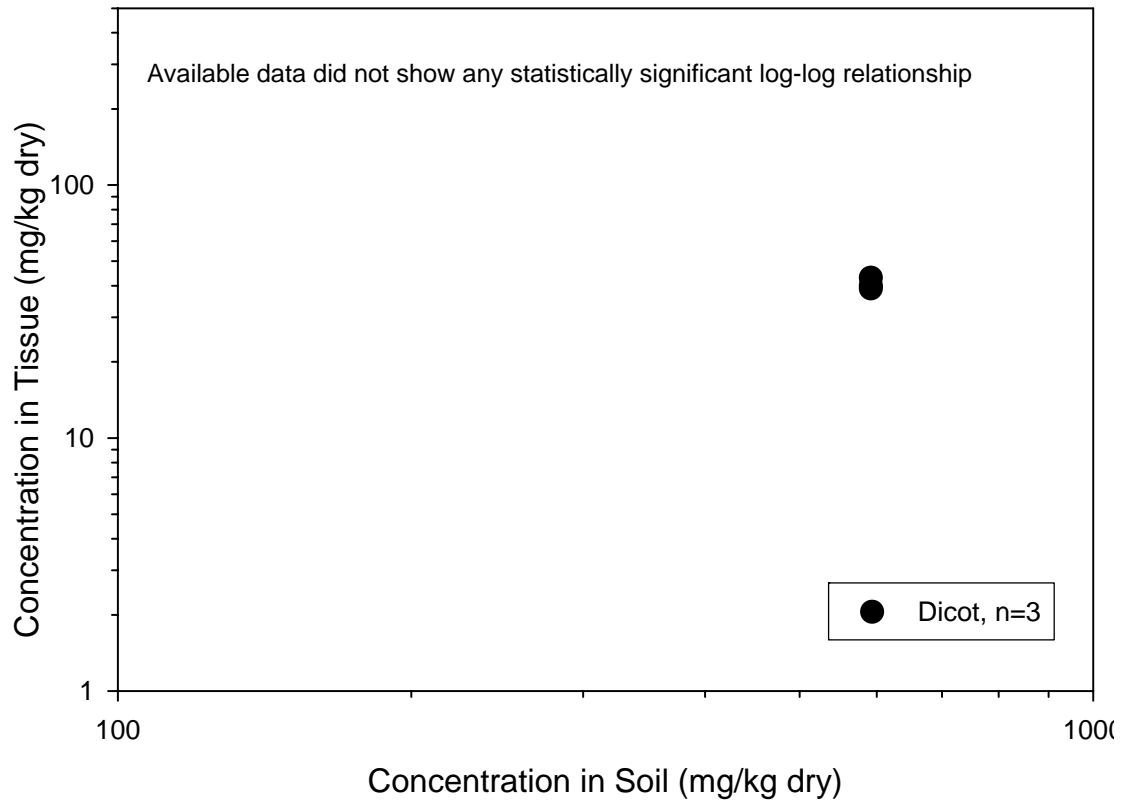


Figure 3.1-10. Perchlorate in soil to Perchlorate in Plant Seed

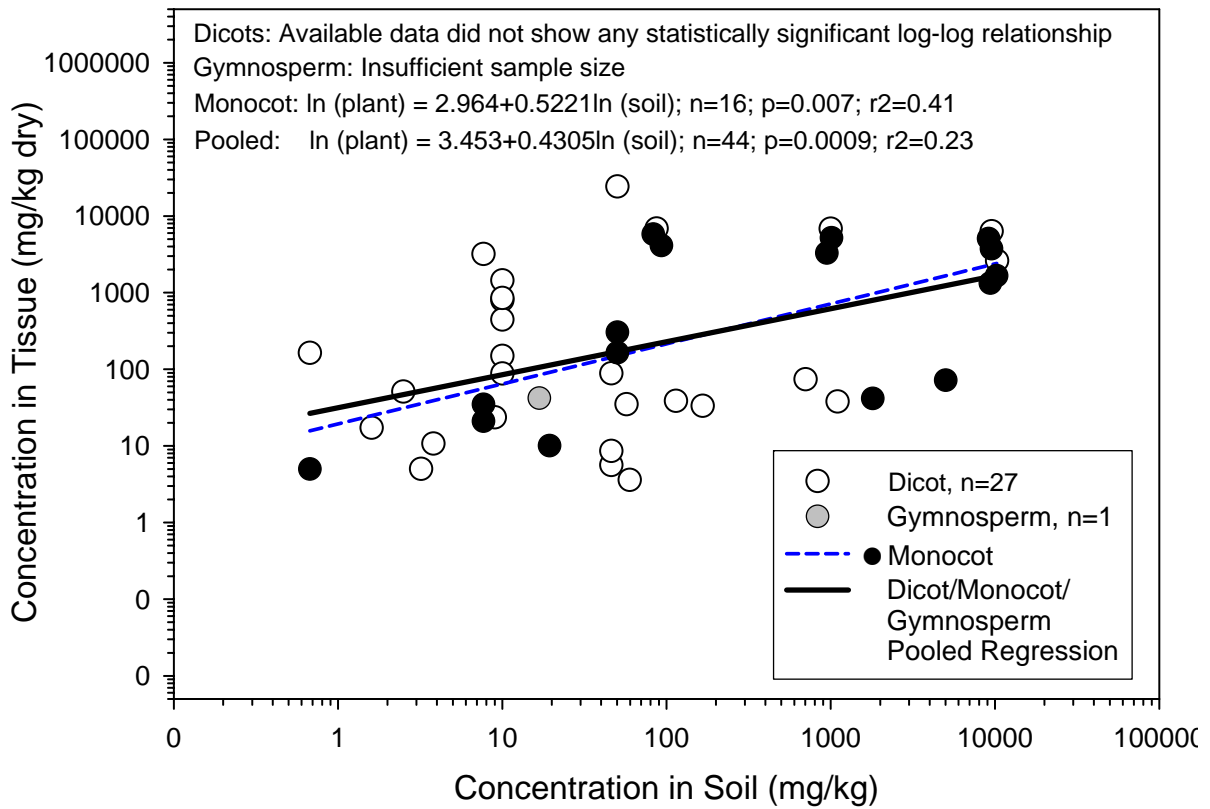


Figure 3.1-11. RDX in Soil to RDX in Plant Foliage

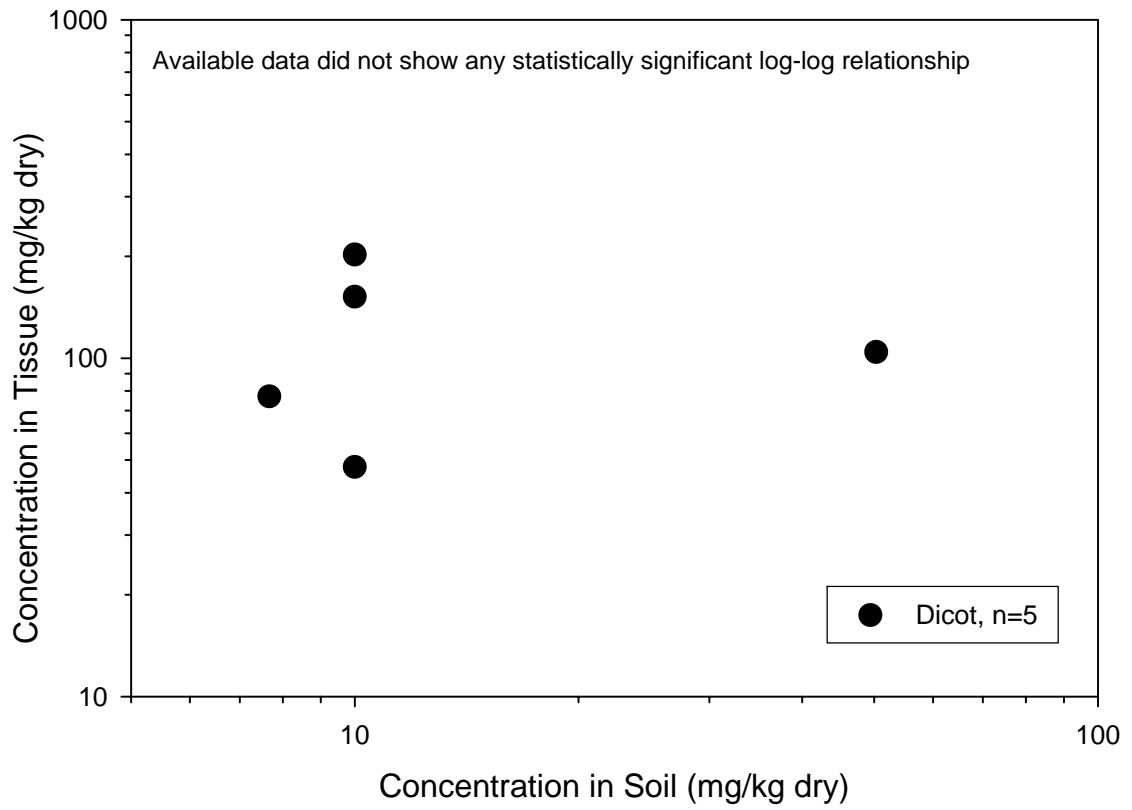


Figure 3.1-12. RDX in Soil to RDX in Fruit

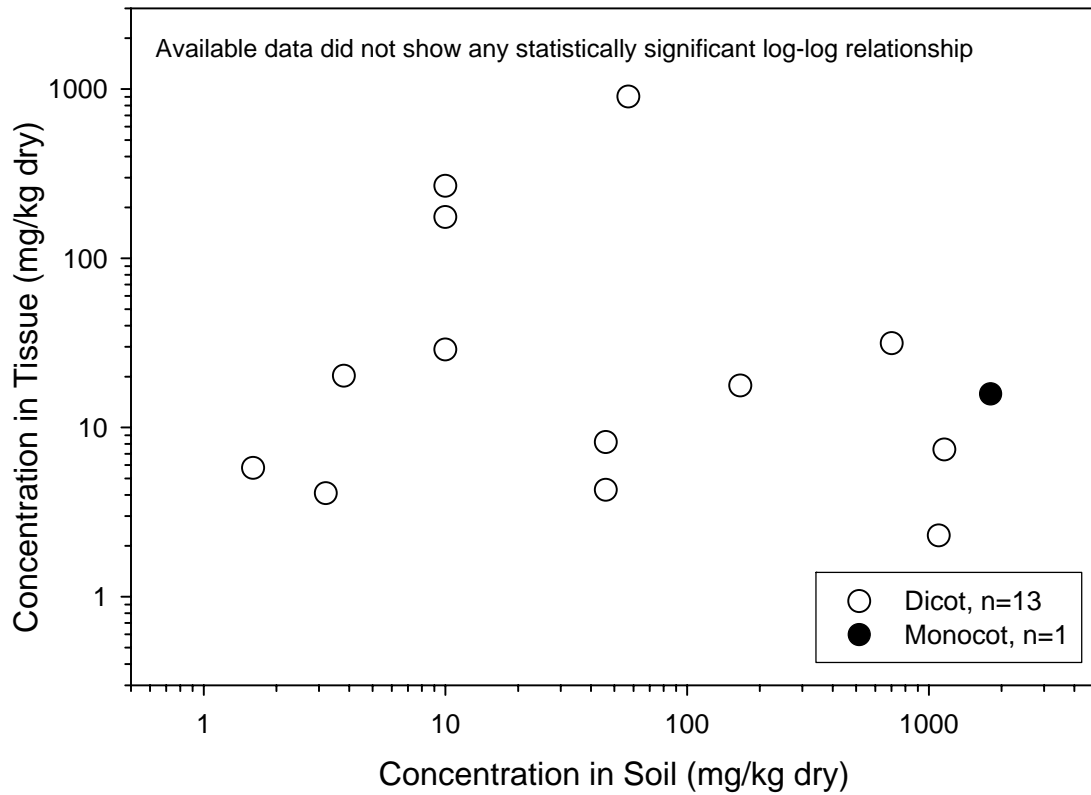


Figure 3.1-13. RDX in Soil to RDX in Plant Root

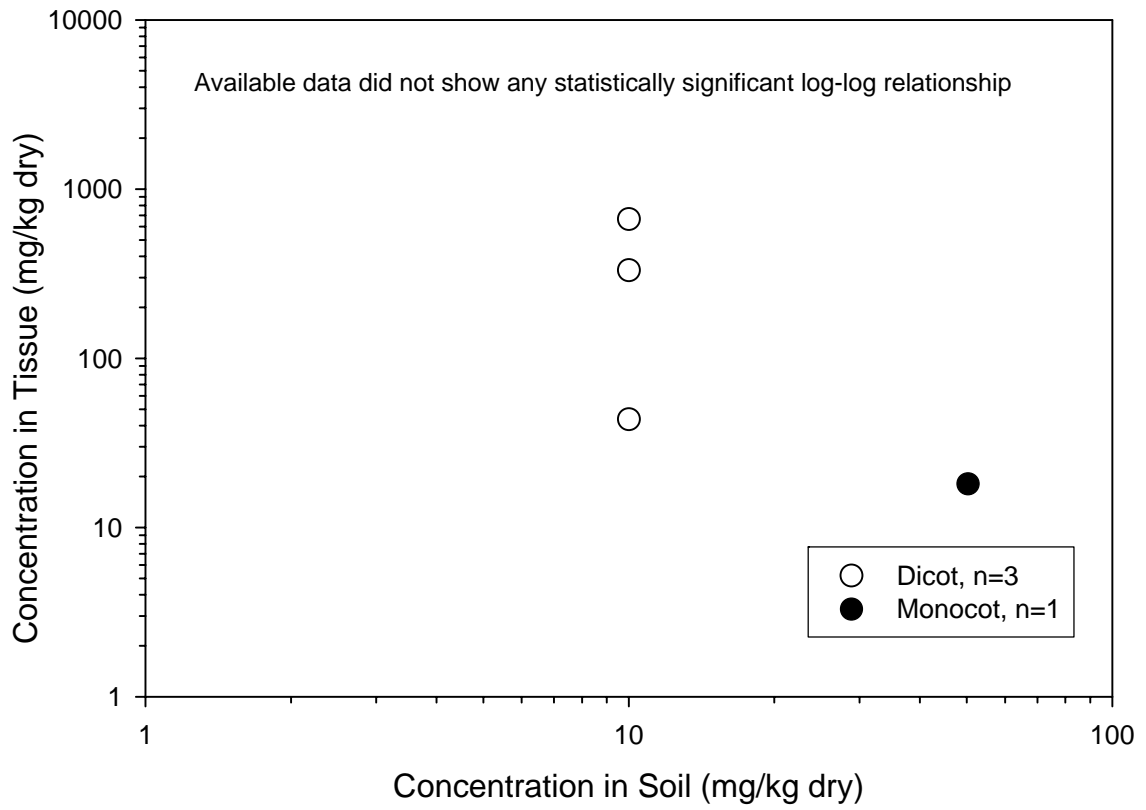


Figure 3.1-14. RDX in Soil to RDX in Plant Seed

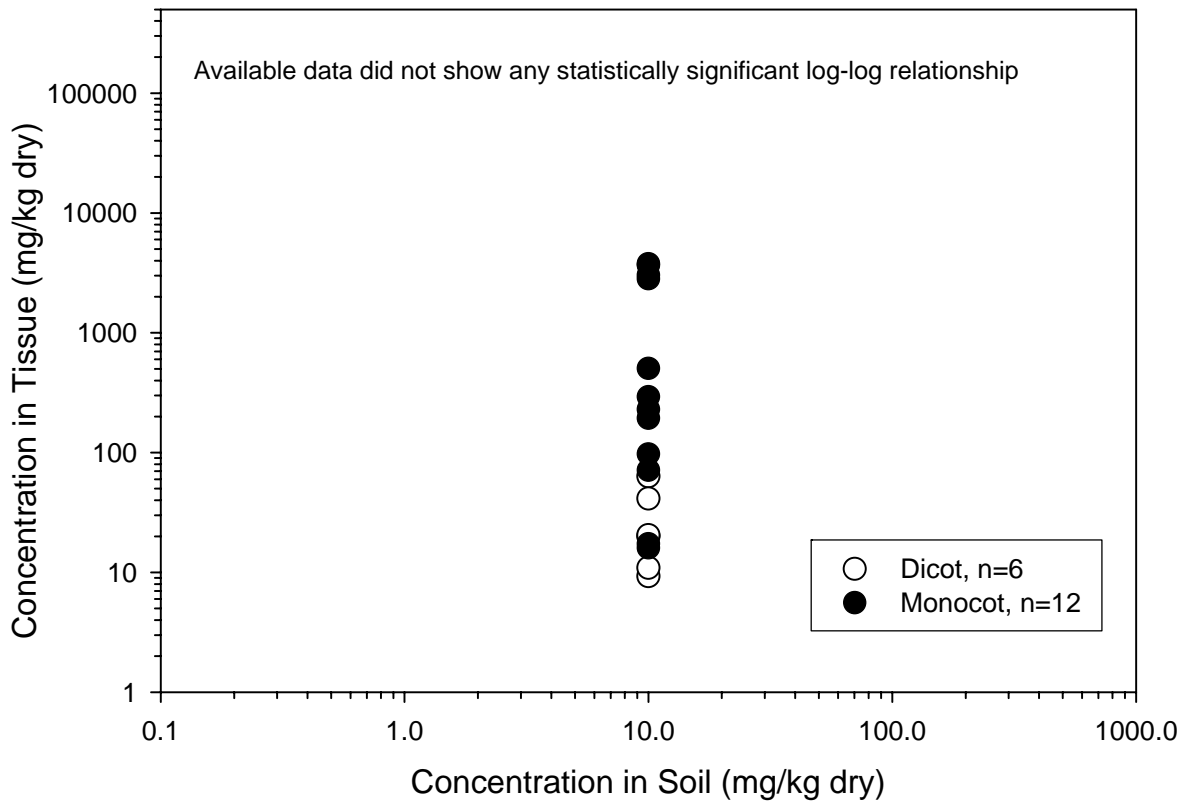


Figure 3.1-15. Tetryl in Soil to Tetryl in Plant Foliage

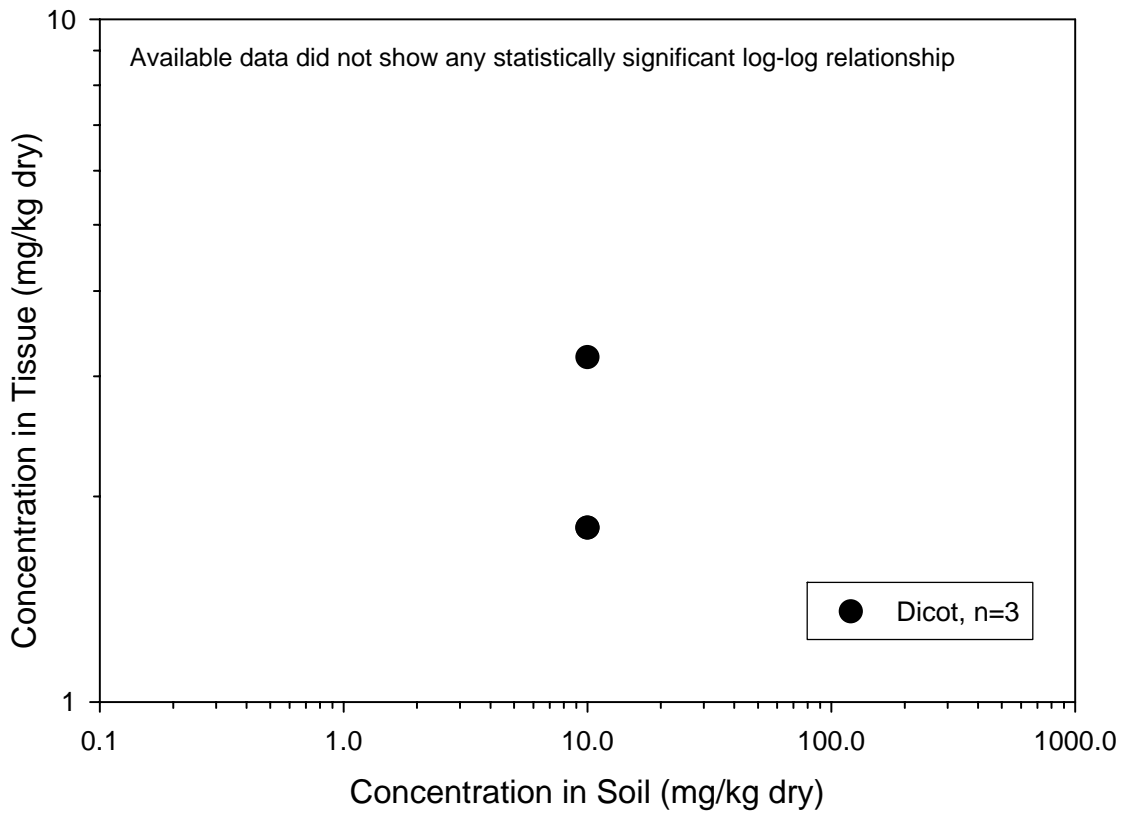


Figure 3.1-16. Tetryl in Soil to Tetryl in Fruits

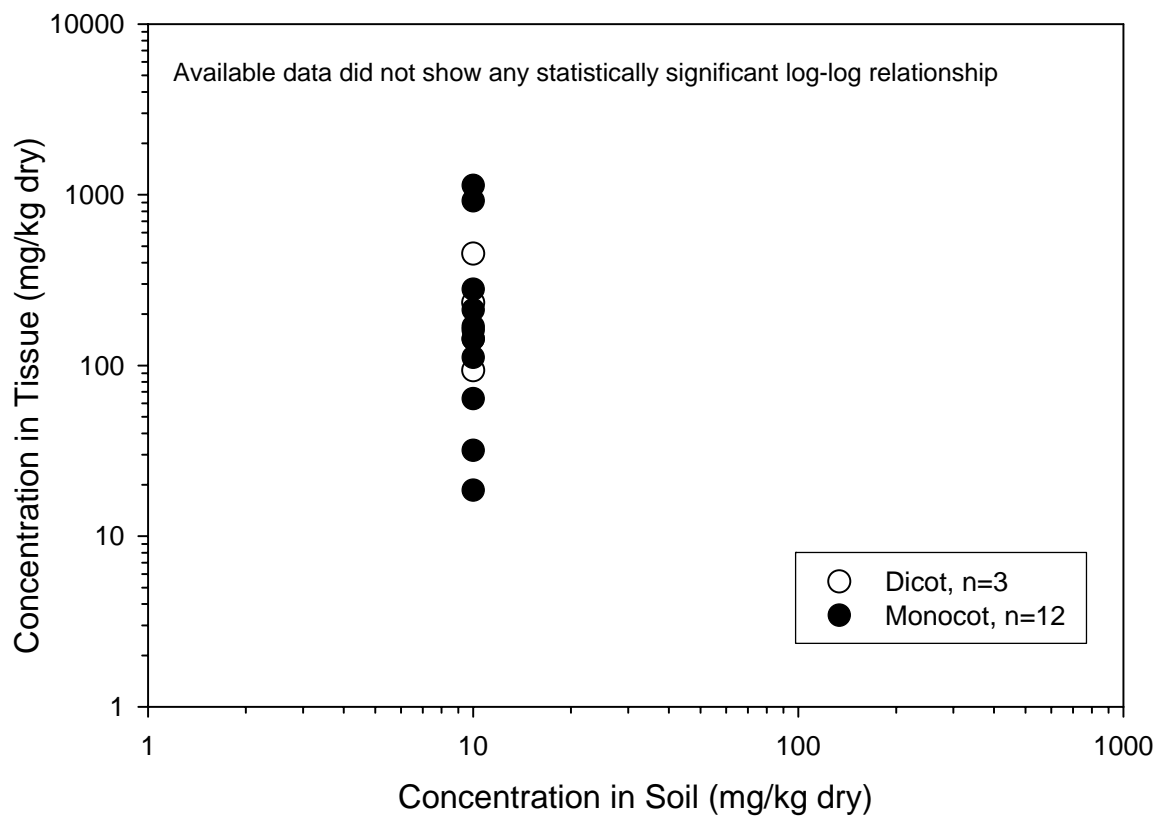


Figure 2.1-17. Tetryl in Soil to Tetryl in Plant Root

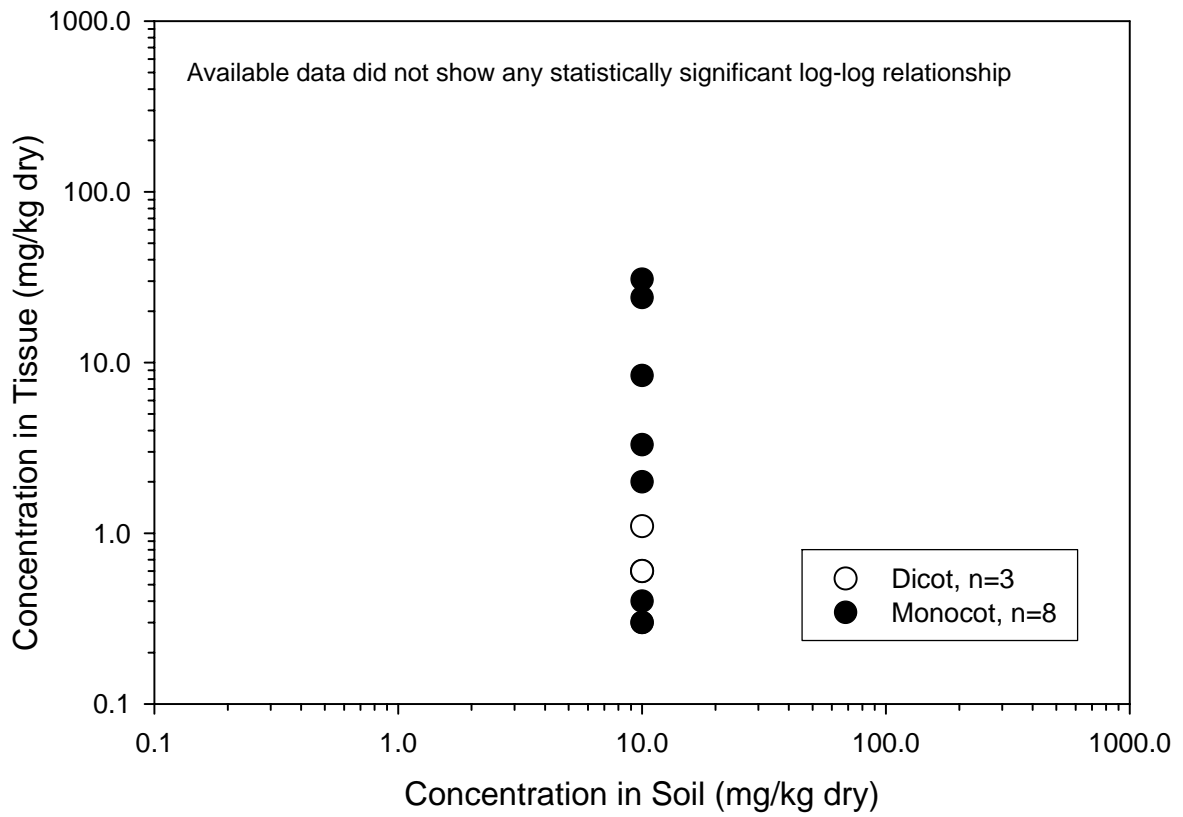


Figure 3.1-18. Tetryl in Soil to Tetryl in Plant Seed

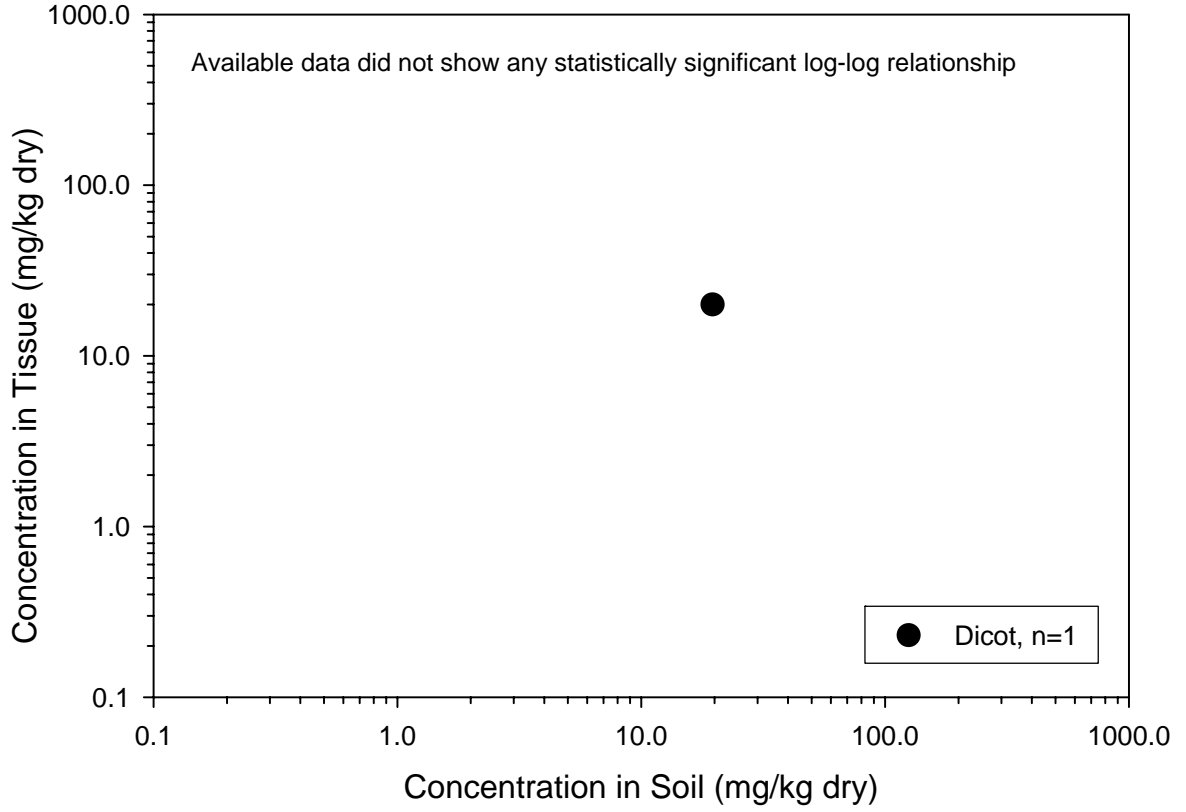


Figure 3.1-19. TNB in Soil to TNB in Plant Foliage

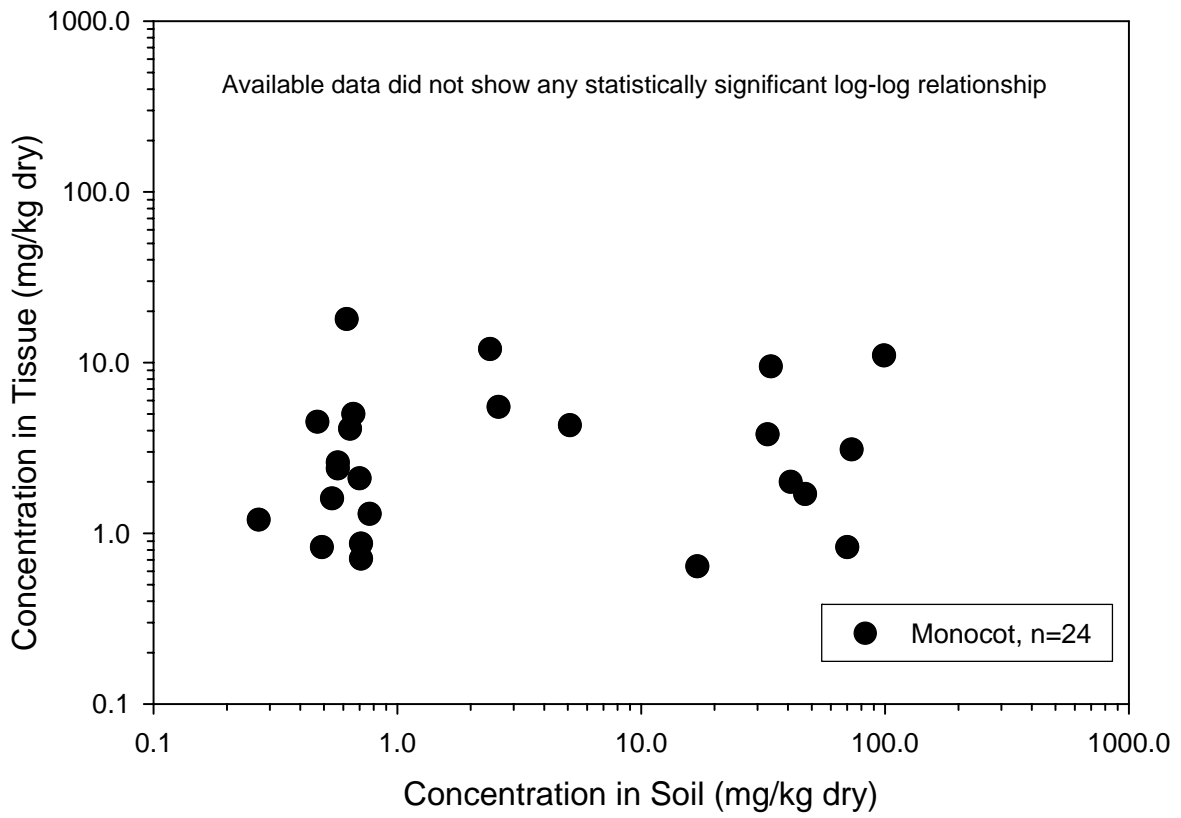


Figure 3.1-20. TNT in Soil to 2-ADNT in Plant Foliage

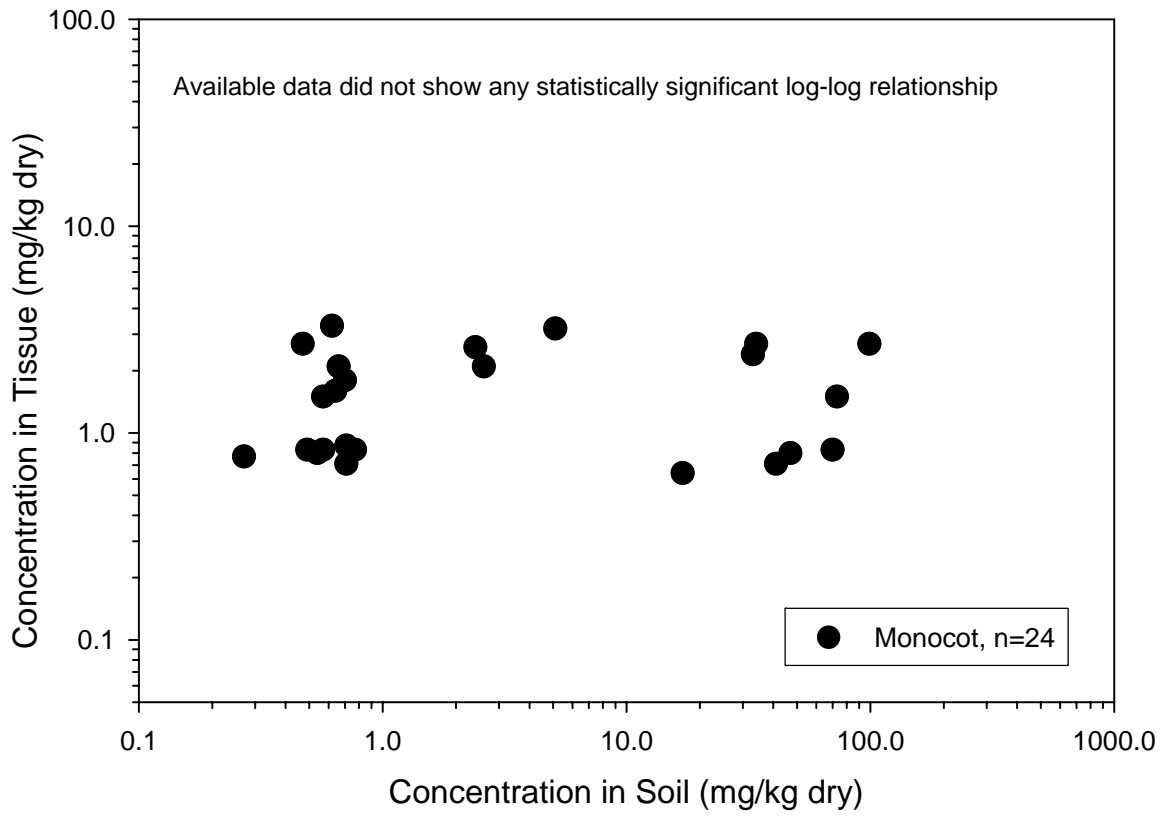


Figure 3.1-21. TNT in Soil to 4-ADNT in Plant Foliage

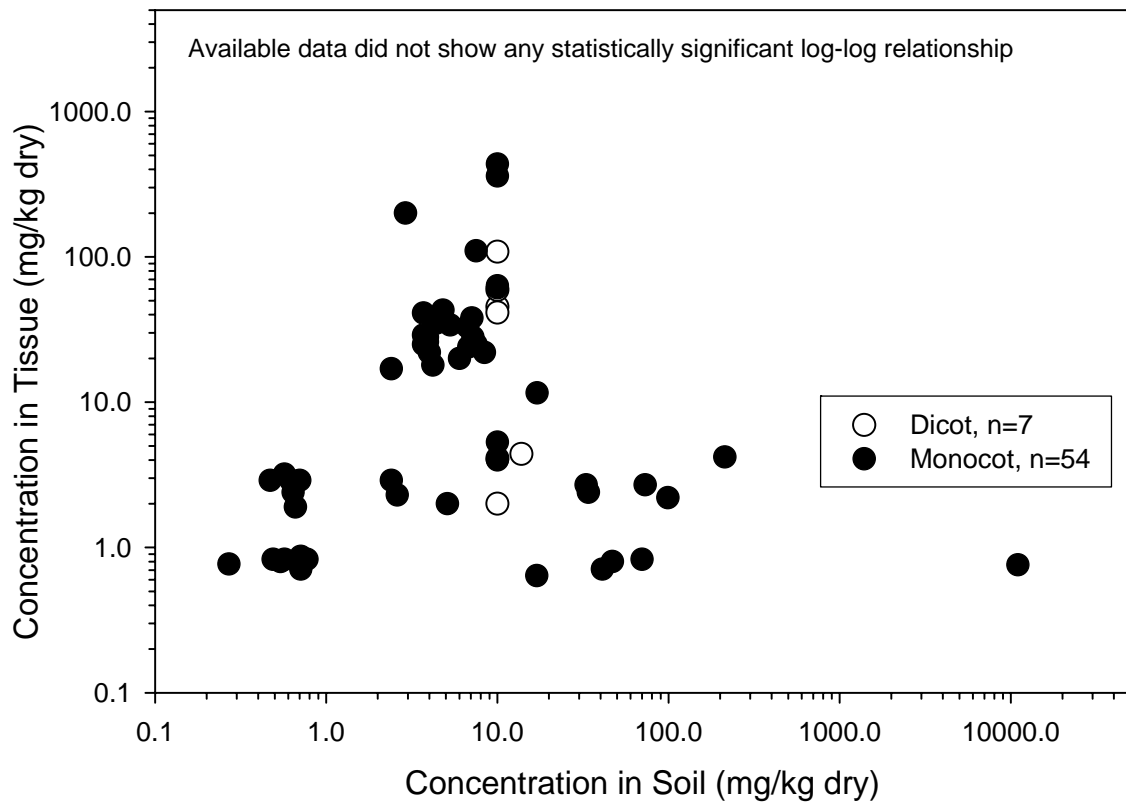


Figure 3.1-22. TNT in Soil to TNT in Plant Foliage

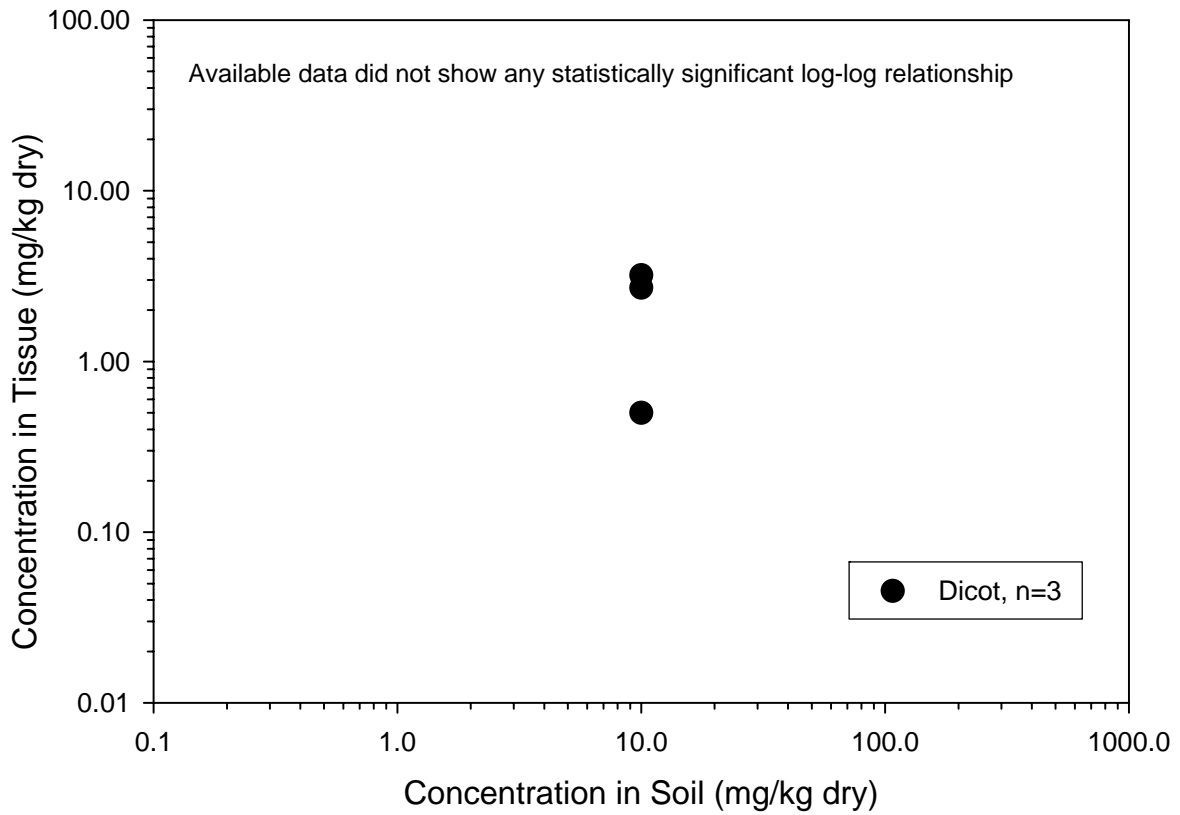


Figure 3.1-23. TNT in Soil to TNT in Fruit

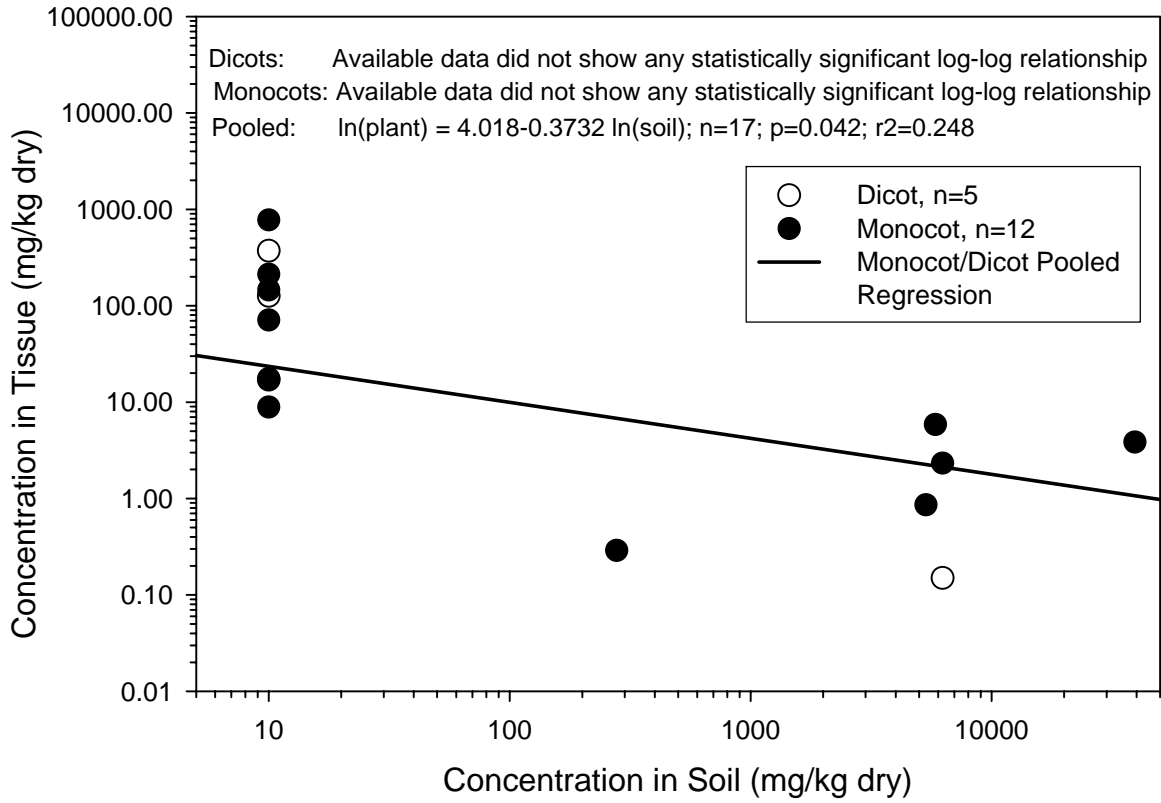


Figure 3.1-24. TNT in Soil to TNT in Plant Root

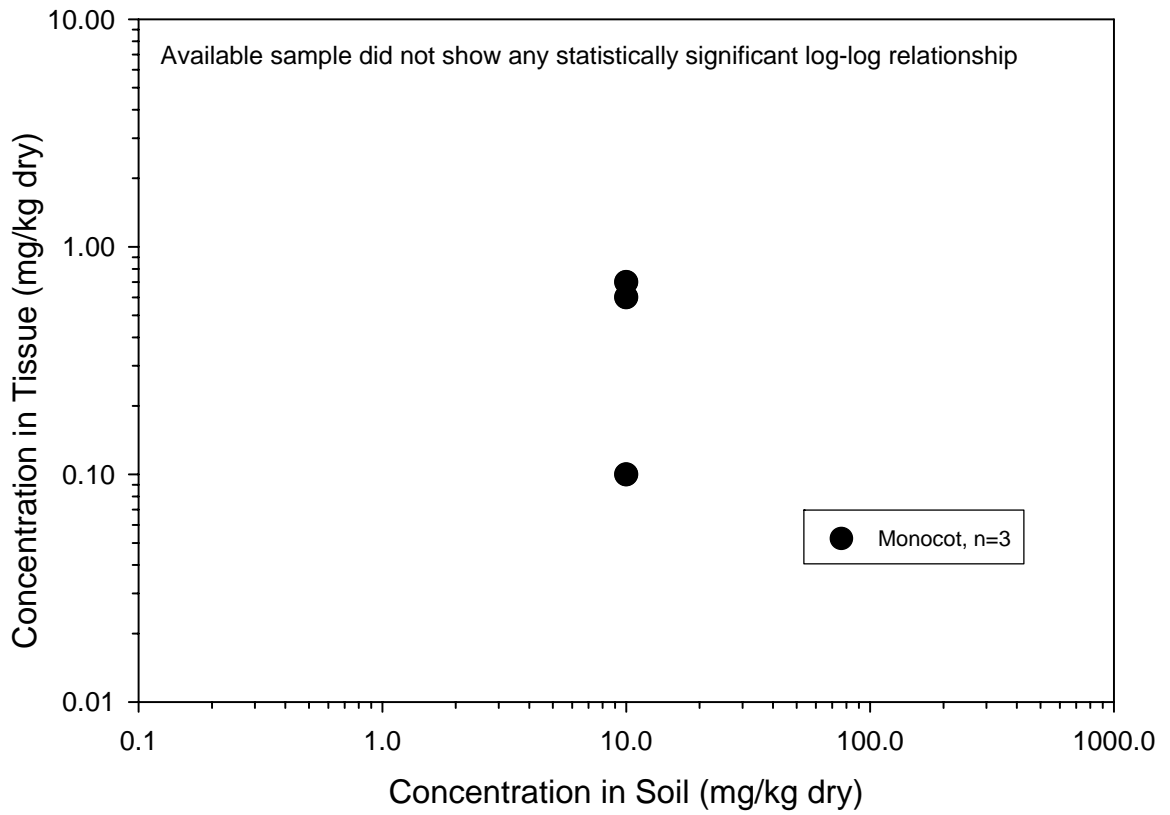


Figure 3.1-25. TNT+2-ADNT in soil to 2-ADNT in Plant Foliage

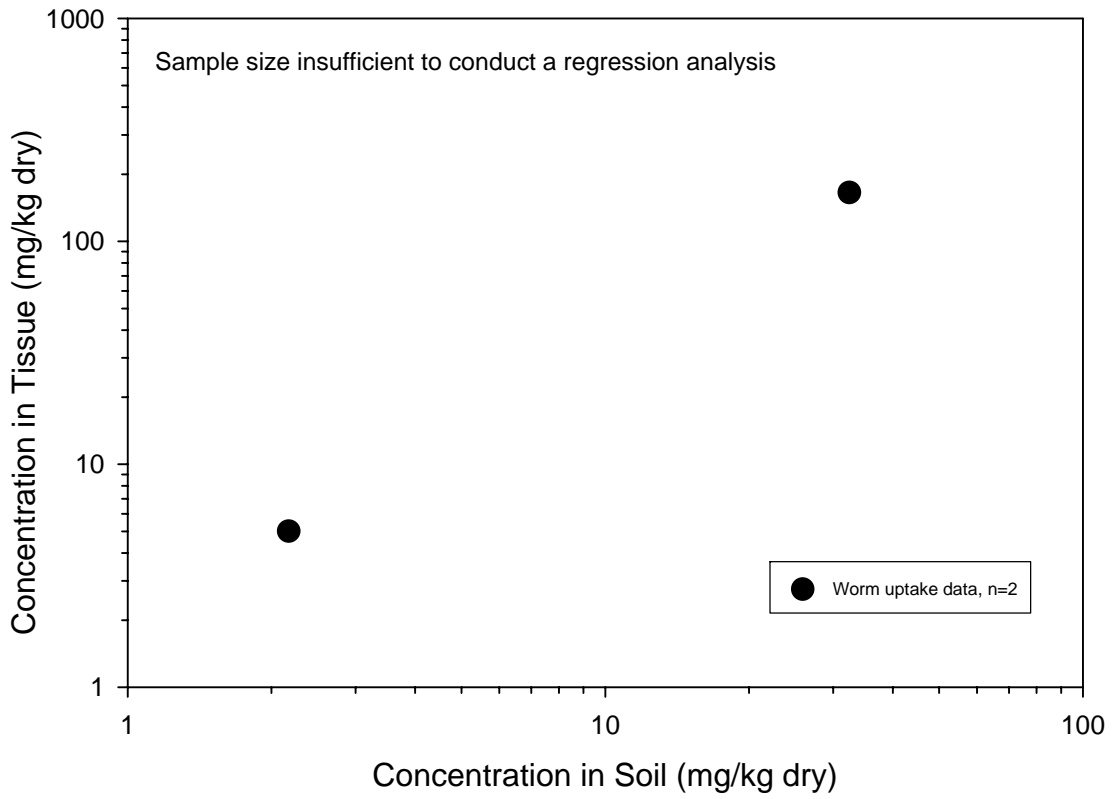


Figure 3.2-1. 2,4-DANT in Soil to 2,4-DANT in Earthworm (whole-body)

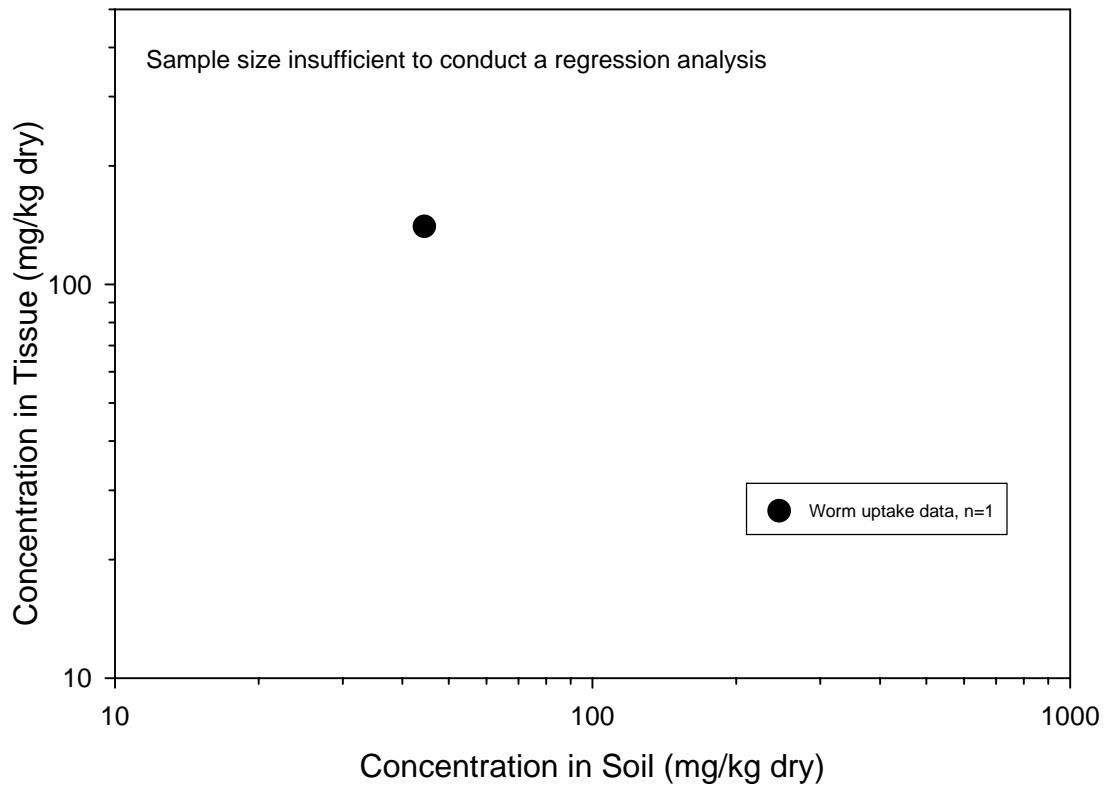


Figure 3.2-2. 2,6-DANT in Soil to 2,6-DANT in Earthworm (whole-body)

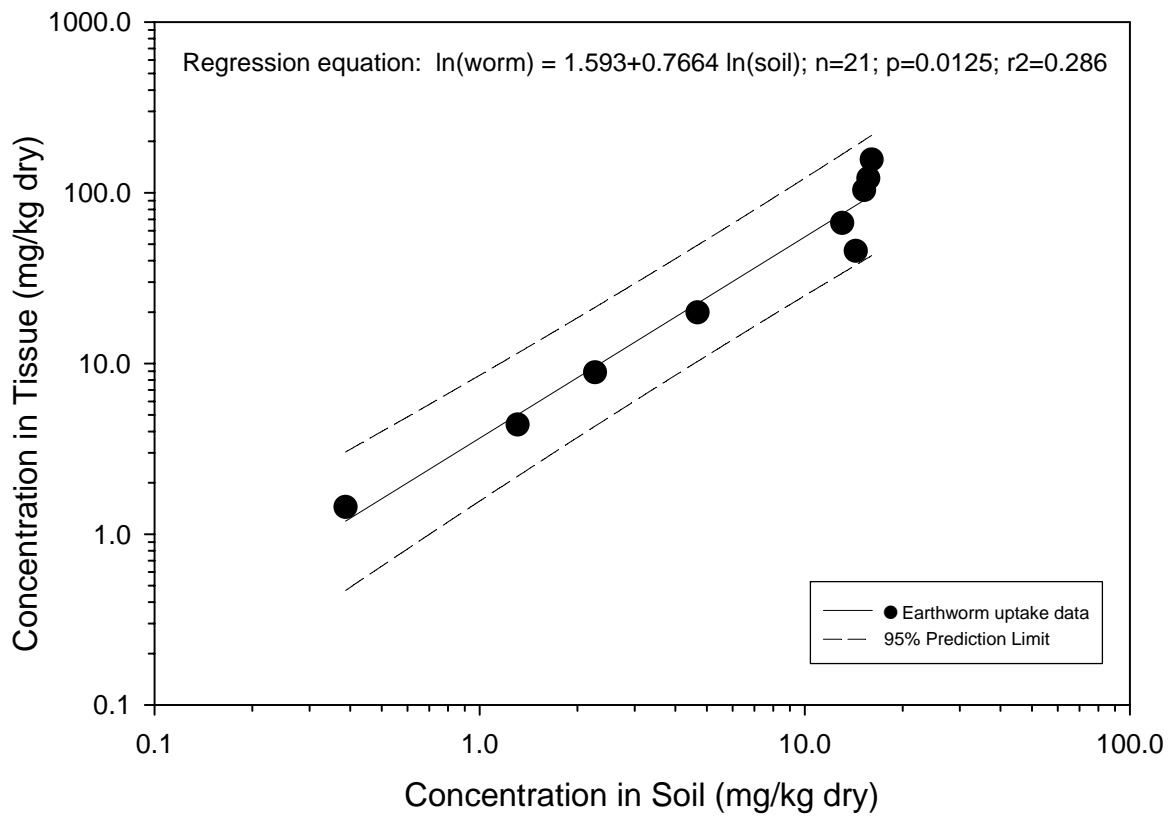


Figure 3.2-3. 2-ADNT in Soil to 2-ADNT in Earthworm (whole-body)

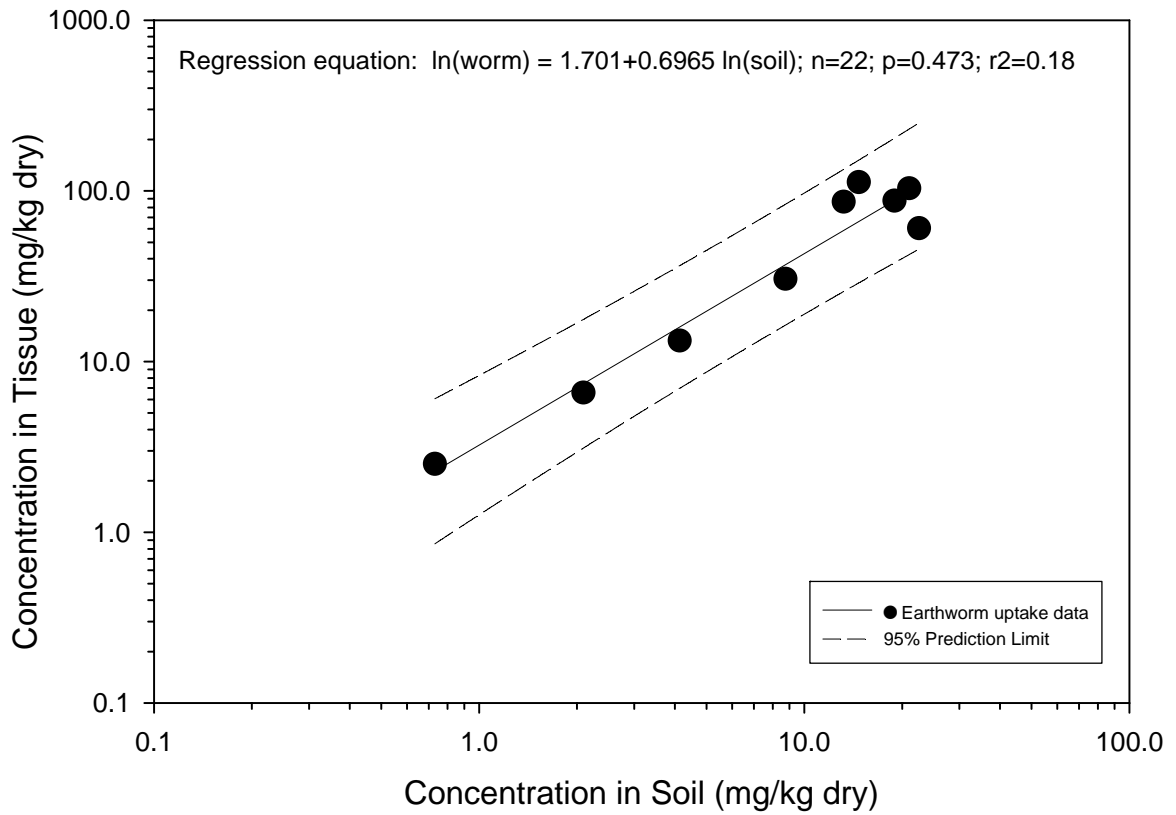


Figure 3.2-4. 4-ADNT in Soil to 4-ADNT in Earthworm (whole-body)

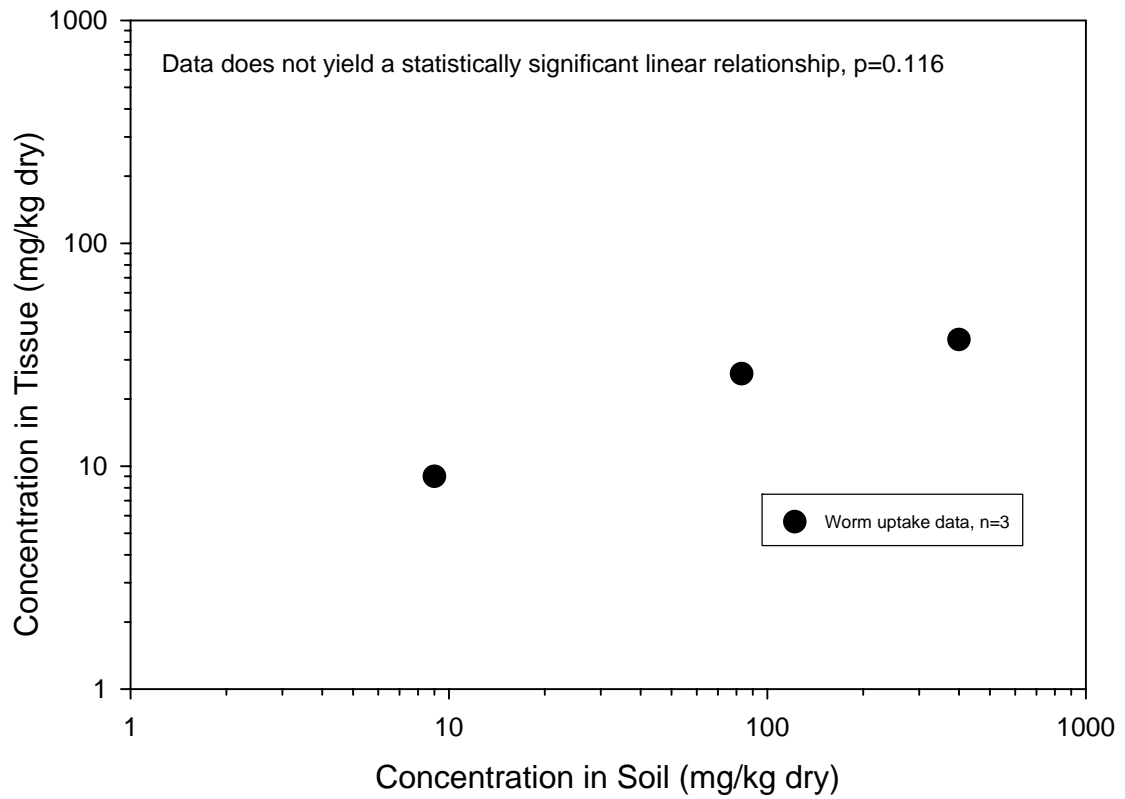


Figure 3.2-5. 2,4-DANT in Soil to 2,4-DANT in Earthworm (whole-body)

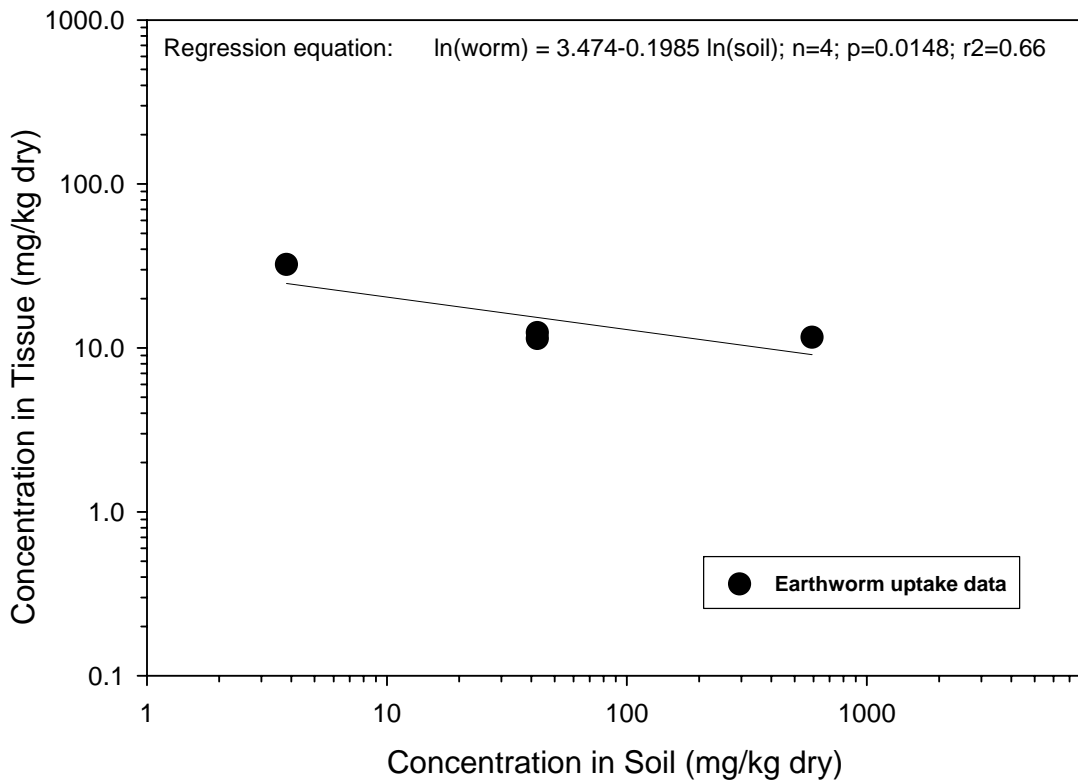


Figure 3.2-6. Perchlorate in Soil to Perchlorate in Earthworm (whole-body)

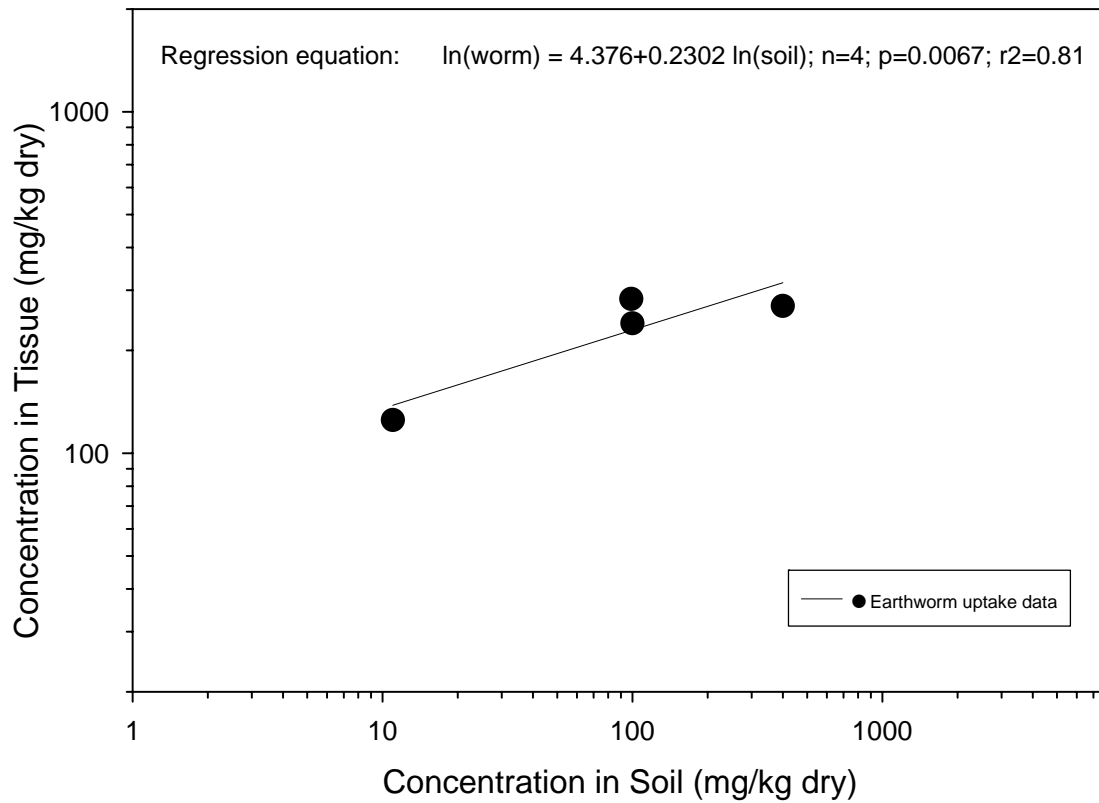


Figure 3.2-7. RDX in Soil to RDX in Earthworm (whole-body)

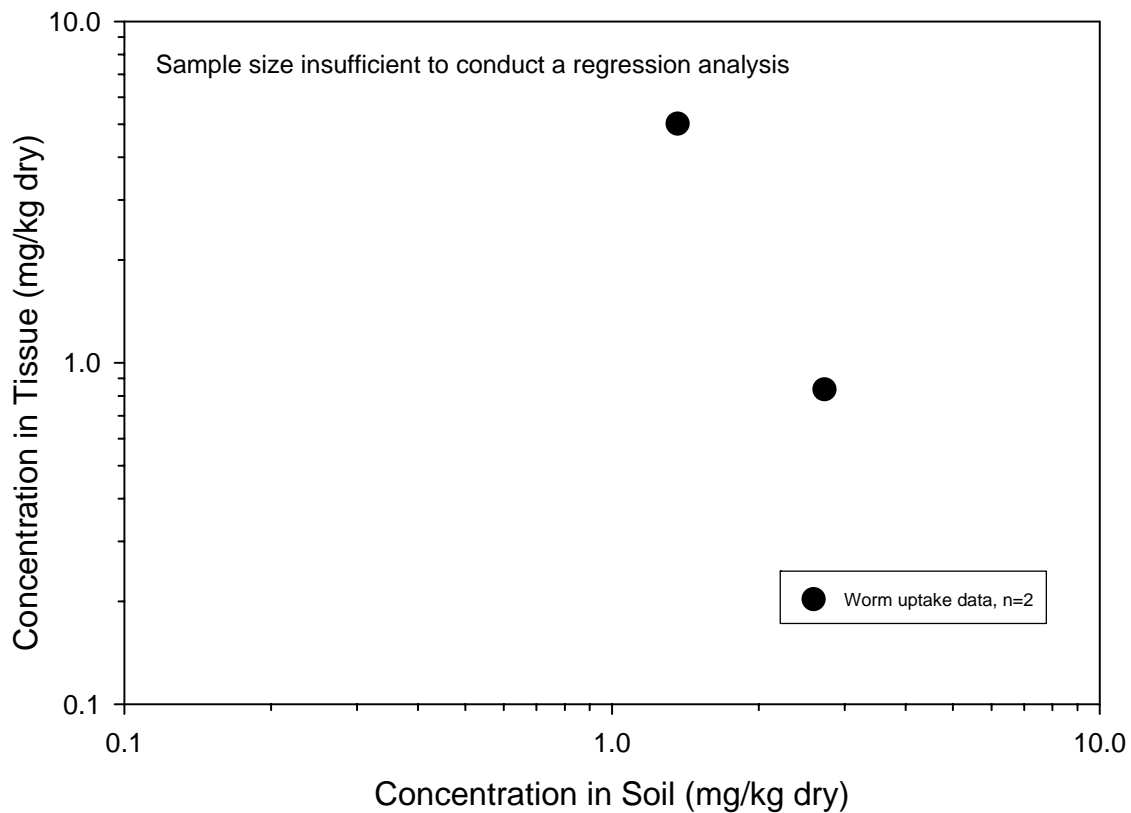


Figure 3.2-8. TNT in Soil to 2,4-DANT in Earthworm (whole-body)

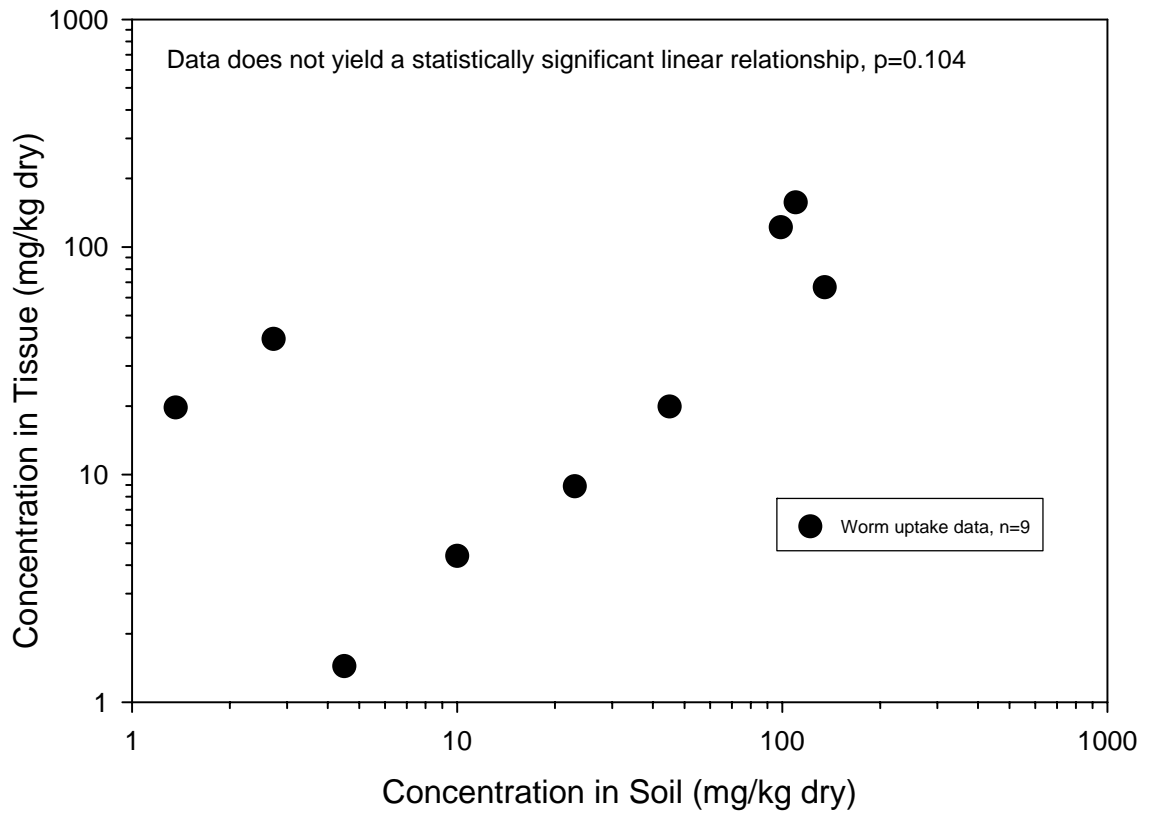


Figure 3.2-9. TNT in Soil to 2-ADNT in Earthworm (whole-body)

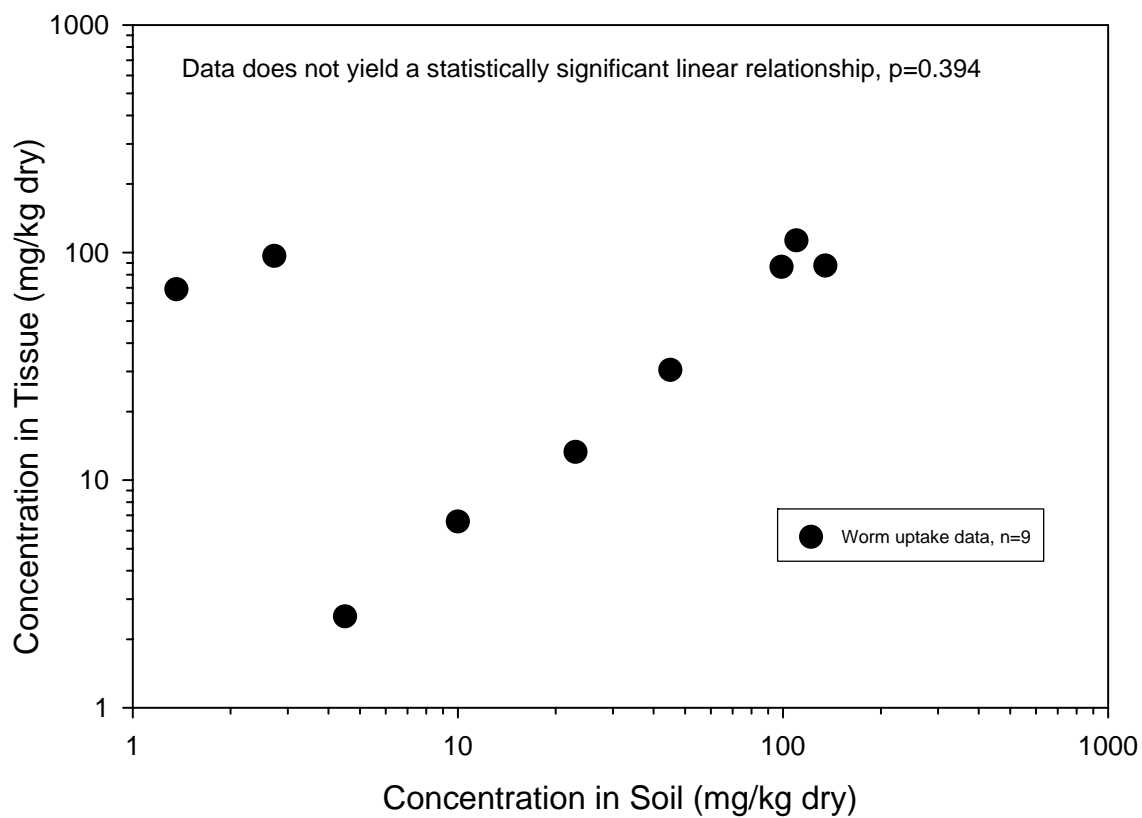


Figure 3.2-10. TNT in Soil to 4-ADNT in Earthworm (whole-body)

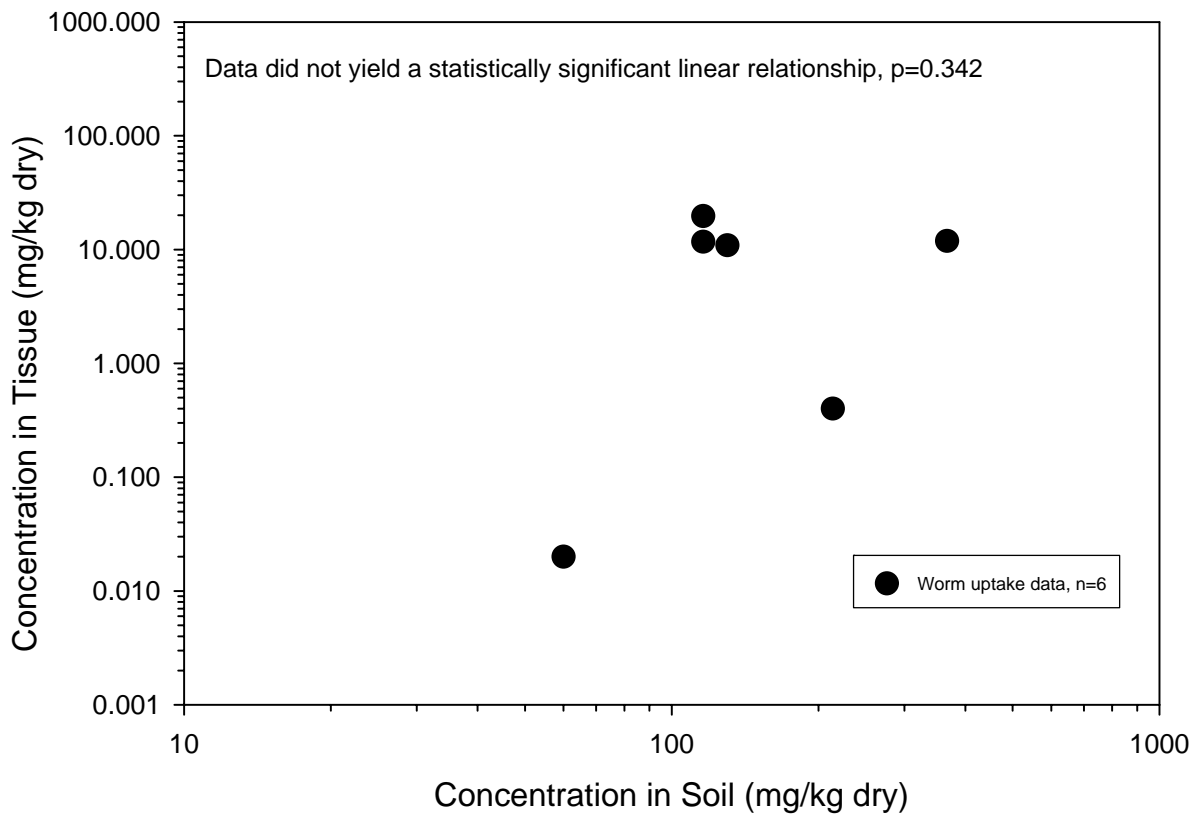


Figure 3.2-11. TNT in Soil to TNT in Earthworm (whole-body)

Appendix A-1
Plant Bioaccumulation (Uptake) Database

Appendix A-1. Plant Bioaccumulation (Uptake) Database

Record NO	Parent Analyte	Analyte in Tissue	Tissue	Plant Species Characteristics								Experimental Condition								Soil Characteristics																				
				Monocot / Dicot	Tissue as Reported	Common Name	Class	Order	Family	Genus	Species	Location	Sub- location	Lab / Field	Duration	Duration Units	Soil Texture	EC	Soil_Ca	Soil_Mg	Nitrate	Bulk_Dens	Field_Capac	Tot_N%	K mg/kg	%Sand	%Silt	%Clay	p	pH	%OM	%OC	CEC							
90	4ADNT	4ADNT	Root	Dicot	Root	Common Teasel	Magnoliopsida	Dipsacales	Dipsacaceae	Dipsacus	<i>Dipsacus sylvestris</i>	Joliet Army Am 7	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--					
91	4ADNT	4ADNT	Root	Dicot	Root	Common Teasel	Magnoliopsida	Dipsacales	Dipsacaceae	Dipsacus	<i>Dipsacus sylvestris</i>	Joliet Army Am 6	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--						
92	4ADNT	4ADNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 8	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--					
93	4ADNT	4ADNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 7	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--				
94	4ADNT	4ADNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 14	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--				
95	4ADNT	4ADNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 13	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--			
96	4ADNT	4ADNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 9	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--			
97	4ADNT	4ADNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 6	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--			
98	4ADNT	4ADNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 12	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
99	HMX	HMX	Foliage	Dicot	Shoot	Alfalfa	Magnoliopsida	Fabales	Fabaceae	Medicago	<i>Medicago sativa</i>	Lab	--	Lab	16	Days	Sandy loam	--	--	--	--	--	--	--	71	18	11	--	5	--	1.3	--	4.27	--	--					
100	HMX	HMX	Foliage	Dicot	Shoot	Alfalfa	Magnoliopsida	Fabales	Fabaceae	Medicago	<i>Medicago sativa</i>	Lab	--	Lab	16	Days	Sandy loam	--	--	--	--	--	--	--	71	18	11	--	5	--	1.3	--	4.27	--	--	--				
101	HMX	HMX	Foliage	Dicot	Shoot	Alfalfa	Magnoliopsida	Fabales	Fabaceae	Medicago	<i>Medicago sativa</i>	Lab	--	Lab	28	Days	Sandy loam	--	--	--	--	--	--	--	71	18	11	--	5	--	1.3	--	4.27	--	--	--	--			
102	HMX	HMX	Foliage	Dicot	Shoot	Alfalfa	Magnoliopsida	Fabales	Fabaceae	Medicago	<i>Medicago sativa</i>	Lab	--	Lab	28	Days	Sandy loam	--	--	--	--	--	--	--	71	18	11	--	5	--	1.3	--	4.27	--	--	--	--	--		
103	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S1W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--			
104	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S1W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--			
105	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S1W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--			
106	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S1W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--	--		
107	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S1W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--	--		
108	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S3W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--	--		
109	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S3W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--	--		
110	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S3W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--	--		
111	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S3W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--	--		
112	HMX	HMX	Foliage	Dicot	Shoot	Lettuce	Magnoliopsida	Asterales	Asteraceae	Lactuca	<i>Lactuca sativa</i>	Lab	S3W0	Lab	45	Days	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--	--	--	--	--	--	--	
113	HMX	HMX	Foliage	Dicot	Leaf	Alfalfa	Magnoliopsida	Fabales	Fabaceae	Medicago	<i>Medicago sativa</i>	Wainwright Firi	--	Lab	77	Days	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--		
114	HMX	HMX	Foliage	Dicot	Leaf	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Wainwright Firi	--	Lab	77	Days	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	
115	HMX	HMX	Foliage	Dicot	Leaf	Canola	Magnoliopsida	Capparales	Brassicaceae	Brassica	<i>Brassica rapa</i>	Wainwright Firi	--	Lab	77	Days	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
116	HMX	HMX	Foliage	Dicot	Leaf	Wild Bergamot	Magnoliopsida	Lamiales	Lamiaceae	Monarda	<i>Monarda fistulosa</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
117	HMX	HMX	Foliage	Dicot	Leaf	Wild Bergamot	Magnoliopsida	Lamiales	Lamiaceae	Monarda	<i>Monarda fistulosa</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
118	HMX	HMX	Foliage	Dicot	Leaf	Wild Bergamot	Magnoliopsida	Lamiales	Lamiaceae	Monarda	<i>Monarda fistulosa</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
119	HMX	HMX	Foliage	Dicot	Leaf	Blueberry	Magnoliopsida	Ericales	Ericaceae	Vaccinium	<i>Vaccinium sp.</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
120	HMX	HMX	Foliage	Dicot	Leaf	Blueberry	Magnoliopsida	Ericales	Ericaceae	Vaccinium	<i>Vaccinium sp.</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
121	HMX	HMX	Foliage	Dicot	Leaf	Blueberry	Magnoliopsida	Ericales	Ericaceae	Vaccinium	<i>Vaccinium sp.</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
122	HMX	HMX	Foliage	Dicot	Leaf	Anemone Sp.	Magnoliopsida	Ranunculales	Ranunculaceae	Anemone	<i>Anemone sp.</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
123	HMX	HMX	Foliage	Dicot	Leaf	Anemone Sp.	Magnoliopsida	Ranunculales	Ranunculaceae	Anemone	<i>Anemone sp.</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
124	HMX	HMX	Foliage	Dicot	Leaf	Anemone Sp.	Magnoliopsida	Ranunculales	Ranunculaceae	Anemone	<i>Anemone sp.</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
125	HMX	HMX	Foliage	Dicot	Leaf	Common Thistle	Magnoliopsida	Asterales	Asteraceae	Cirsium	<i>Cirsium vulgare</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
126	HMX	HMX	Foliage	Dicot	Leaf	Common Thistle	Magnoliopsida	Asterales	Asteraceae	Cirsium	<i>Cirsium vulgare</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
127	HMX	HMX	Foliage	Dicot	Leaf	Common Thistle	Magnoliopsida	Asterales	Asteraceae	Cirsium	<i>Cirsium vulgare</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
128	HMX	HMX	Foliage	Dicot	Leaf	Wax Berry	Magnoliopsida	Dipsacales	Caprifoliaceae	Symphoricarpos	<i>Symphoricarpos albus</i>	Wainwright Firi	--	Field	Resident	--	--	--	--	--	--	--	--	--	59	25	16	< 2.5	7.52	--	2.1	18	--	--	--	--	--	--	--	--
129	HMX	HMX	Foliage	Dicot	Leaf	Wax Berry	Magnoliopsida	Dipsacales	Caprifoliaceae	Symphoricarpos	<i>Symphoricarpos albus</i>	Wainwright Firi	--	Field	Resident																									

Appendix A-1. Plant Bioaccumulation (Uptake) Database

Record NO	Plant Species Characteristics											Experimental Condition						Soil Characteristics																		
	Parent Analyte	Analyte in Tissue	Tissue	Monocot / Dicot	Tissue as Reported	Common Name	Class	Order	Family	Genus	Species	Location	Sub-location	Lab / Field	Duration	Units	Soil Texture	EC	Soil_Ca	Soil_Mg	Nitrate	Bulk_Dens	Field_Capac	Tot_N%	K mg/kg	%Sand	%Silt	%Clay	p	pH	%OM	%OC	CEC			
272	RDX	RDX	Foliage	Monocot	Leaf/Stem	Reed Canary Grass	Liliopsida	Cyperales	Poaceae	Phalaris	<i>Phalaris arundinacea</i>	Iowa Army Am 12		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
273	RDX	RDX	Foliage	Monocot	Leaf/Stem	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Iowa Army Am 105		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
274	RDX	RDX	Fruit	Dicot	Fruit	Tomato	Magnoliopsida	Scrophulariales	Solanaceae	Lycopersicon	<i>Lycopersicon esculentum</i>	Lab	S1W0	Lab	50-85	Days	--	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--			
275	RDX	RDX	Fruit	Dicot	Fruit	Tomato	Magnoliopsida	Scrophulariales	Solanaceae	Lycopersicon	<i>Lycopersicon esculentum</i>	Lab	S3W0	Lab	50-85	Days	--	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--			
276	RDX	RDX	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--			
277	RDX	RDX	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--			
278	RDX	RDX	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--		
279	RDX	RDX	Root	Dicot	Root	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
280	RDX	RDX	Root	Dicot	Root	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
281	RDX	RDX	Root	Dicot	Root	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--		
282	RDX	RDX	Root	Dicot	Root	Arrowhead	Magnoliopsida	Alismatales	Alismataceae	Sagittaria	<i>Sagittaria calycina</i>	Iowa Army Am 8		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
283	RDX	RDX	Root	Dicot	Root	Goldenrod	Magnoliopsida	Asterales	Asteraceae	Solidago	<i>Solidago canadensis</i>	Iowa Army Am 18		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	
284	RDX	RDX	Root	Dicot	Root	Ragweed	Magnoliopsida	Asterales	Asteraceae	Ambrosia	<i>Ambrosia artemisiifolia</i>	Iowa Army Am 21		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	
285	RDX	RDX	Root	Dicot	Root	Smartweed	Magnoliopsida	Polygonales	Polygonaceae	Polygonum	<i>Polygonum sp.</i>	Iowa Army Am 104		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--
286	RDX	RDX	Root	Dicot	Root	Pigweed	Magnoliopsida	Caryophyllales	Amaranthaceae	Amaranthus	<i>Amaranthus sp.</i>	Iowa Army Am 106		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	
287	RDX	RDX	Root	Dicot	Root	Ragweed	Magnoliopsida	Asterales	Asteraceae	Ambrosia	<i>Ambrosia artemisiifolia</i>	Iowa Army Am 108		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--
288	RDX	RDX	Root	Dicot	Root	Goldenrod	Magnoliopsida	Asterales	Asteraceae	Solidago	<i>Solidago canadensis</i>	Iowa Army Am 109		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--
289	RDX	RDX	Root	Dicot	Root	Black Locust	Magnoliopsida	Fabales	Fabaceae	Robinia	<i>Robinia pseudo-acacia</i>	Iowa Army Am 111		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	
290	RDX	RDX	Root	Dicot	Root	Common Milkweed	Magnoliopsida	Gentianales	Asclepiadaceae	Asclepias	<i>Asclepias syriaca</i>	Iowa Army Am 112-1		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	
291	RDX	RDX	Root	Dicot	Root	Sunflower	Magnoliopsida	Asterales	Asteraceae	Helianthus	<i>Helianthus nuttallii</i>	Iowa Army Am 112-3		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	
292	RDX	RDX	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Iowa Army Am 105		Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	
293	RDX	RDX	Seed	Dicot	Seed	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
294	RDX	RDX	Seed	Dicot	Seed	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
295	RDX	RDX	Seed	Dicot	Seed	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--	--	
296	RDX	RDX	Seed	Monocot	Seed	Corn	Liliopsida	Cyperales	Poaceae	Zea	<i>Zea mays</i>	Lab	S3W0	Lab	76-80	Days	--	--	2505	720.5	1339	--	--	28.5	5.14	31.5	46.6	21.9	429	5.65	--	--	--	--		
297	Tetryl	Tetryl	Foliage	Dicot	Leaf	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
298	Tetryl	Tetryl	Foliage	Dicot	Leaf	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
299	Tetryl	Tetryl	Foliage	Dicot	Leaf	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--		
300	Tetryl	Tetryl	Foliage	Dicot	Stem	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
301	Tetryl	Tetryl	Foliage	Dicot	Stem	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
302	Tetryl	Tetryl	Foliage	Dicot	Stem	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--		
303	Tetryl	Tetryl	Foliage	Monocot	Leaf	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
304	Tetryl	Tetryl	Foliage	Monocot	Leaf	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
305	Tetryl	Tetryl	Foliage	Monocot	Leaf	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--		
306	Tetryl	Tetryl	Foliage	Monocot	Leaf	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
307	Tetryl	Tetryl	Foliage	Monocot	Leaf	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
308	Tetryl	Tetryl	Foliage	Monocot	Leaf	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--		
309	Tetryl	Tetryl	Foliage	Monocot	Leaf	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
310	Tetryl	Tetryl	Foliage	Monocot	Leaf	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
311	Tetryl	Tetryl	Foliage	Monocot	Leaf	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--		
312	Tetryl	Tetryl	Foliage	Monocot	Leaf	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
313	Tetryl	Tetryl	Foliage	Monocot	Leaf	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
314	Tetryl	Tetryl	Foliage	Monocot	Leaf	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	--	--		
315	Tetryl	Tetryl	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--	--	--		
316	Tetryl	Tetryl	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	--	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	--	--		
317	Tetryl	Tetryl	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>																									

Appendix A-1. Plant Bioaccumulation (Uptake) Database

Record NO	Plant Species Characteristics											Experimental Condition						Soil Characteristics															
	Parent Analyte	Analyte in Tissue	Tissue	Monocot / Dicot	Tissue as Reported	Common Name	Class	Order	Family	Genus	Species	Location	Sub- location	Lab / Field	Duration	Duration Units	Soil Texture	EC	Soil_Ca	Soil_Mg	Nitrate	Bulk_Dens	Field_Capac	Tot_N%	K mg/kg	%Sand	%Silt	%Clay	p	pH	%OM	%OC	CEC
454	TNT	TNT	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Clay loam	--	--	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--
455	TNT	TNT	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Silt loam	--	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--	
456	TNT	TNT	Fruit	Dicot	Pod	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Sandy loam	--	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--	
457	TNT	TNT	Root	Dicot	Root	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Clay loam	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--		
458	TNT	TNT	Root	Dicot	Root	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Silt loam	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--		
459	TNT	TNT	Root	Dicot	Root	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Sandy loam	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--		
460	TNT	TNT	Root	Dicot	Root	Common Milkweed	Magnoliopsida	Gentianales	Asclepiadaceae	Asclepias	<i>Asclepias syriaca</i>	Joliet Army Am 3	--	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
461	TNT	TNT	Root	Dicot	Root	Common Teasel	Magnoliopsida	Dipsacales	Dipsacaceae	Dipsacus	<i>Dipsacus sylvestris</i>	Joliet Army Am 6	--	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
462	TNT	TNT	Root	Monocot	Root	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	Clay loam	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--		
463	TNT	TNT	Root	Monocot	Root	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	Silt loam	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--		
464	TNT	TNT	Root	Monocot	Root	Wheat	Liliopsida	Cyperales	Poaceae	Triticum	<i>Triticum aestivum</i>	Lab	--	Lab	60	Days	Sandy loam	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--		
465	TNT	TNT	Root	Monocot	Root	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	Clay loam	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--		
466	TNT	TNT	Root	Monocot	Root	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	Silt loam	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--		
467	TNT	TNT	Root	Monocot	Root	Bland Brome	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus hordeaceus</i>	Lab	--	Lab	60	Days	Sandy loam	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--		
468	TNT	TNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 3	--	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
469	TNT	TNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 8	--	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
470	TNT	TNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 13	--	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
471	TNT	TNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 9	--	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
472	TNT	TNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 6	--	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
473	TNT	TNT	Root	Monocot	Root	Smooth Bromegrass	Liliopsida	Cyperales	Poaceae	Bromus	<i>Bromus inermis</i>	Joliet Army Am 12	--	Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--	--		
474	TNT	TNT	Seed	Dicot	Seed	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Clay loam	--	--	--	--	--	--	--	--	--	--	--	7.2	--	--		
475	TNT	TNT	Seed	Dicot	Seed	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Silt loam	--	--	--	--	--	--	--	--	--	--	--	1.7	--	--		
476	TNT	TNT	Seed	Dicot	Seed	Bush Bean	Magnoliopsida	Fabales	Fabaceae	Phaseolus	<i>Phaseolus vulgaris</i>	Lab	--	Lab	60	Days	Sandy loam	--	--	--	--	--	--	--	--	--	--	--	0.5	--	--		

Appendix A-1. Plant Bioaccumulation (Uptake) Database

Results															
Record NO	Soil Conc	Soil Qualifier	Soil Units	Soil Measured Conc	soil_wet_dry?	Water Conc	Water Units	Water Qualifier	Tissue Conc	Tissue Qualifier	Tissue Units	Reported BAF	N	Reference	Notes:
454	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	0.5	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 0.1 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 78
455	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	3.2	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 0.7 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 78
456	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	2.7	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 0.6 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 78
457	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	17.1	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 4.8 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
458	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	126.4	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 35.4 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
459	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	371.4	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 104 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
460	1	Det	mg/kg-dw	M	--	--	--	--	0.6	Det	--	--	--	Zellmer et al. 1995	--
461	6260	Det	mg/kg-dw	M	--	--	--	--	0.15	Det	--	--	--	Zellmer et al. 1995	--
462	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	17.5	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 4.9 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
463	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	146.1	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 40.9 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
464	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	775	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 217 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
465	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	8.9	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 2.5 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
466	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	71.1	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 19.9 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
467	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	212.1	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 59.4 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 72
468	1	Det	mg/kg-dw	M	--	--	--	--	4.5	Det	--	--	--	Zellmer et al. 1995	--
469	278	Det	mg/kg-dw	M	--	--	--	--	0.29	Det	--	--	--	Zellmer et al. 1995	--
470	5340	Det	mg/kg-dw	M	--	--	--	--	0.86	Det	--	--	--	Zellmer et al. 1995	--
471	5840	Det	mg/kg-dw	M	--	--	--	--	5.85	Det	--	--	--	Zellmer et al. 1995	--
472	6260	Det	mg/kg-dw	M	--	--	--	--	2.33	Det	--	--	--	Zellmer et al. 1995	--
473	39350	Det	mg/kg-dw	M	--	--	--	--	3.85	Det	--	--	--	Zellmer et al. 1995	--
474	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	0.1	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 0.1 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 9.3
475	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	0.7	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 0.6 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 9.3
476	10	Det	mg/kg-dw	Nominal	dry-weight	--	--	--	0.6	Det	mg/kg-dry	--	--	Cataldo et al. 1993	Source: Tbl 5, plant wet weight conc is 0.5 / Assumed %WC because plant concentrations are reported as wet weight basis / Authors noted uptake is dependent on soil type (sand > silt >> clay) and plant type / %WC Assumed is: 9.3

Appendix A-2
Earthworm Bioaccumulation (Uptake) Database

Appendix A-2. Soil Invertebrate Bioaccumulation (Uptake) Database

Soil Invertebrate Species Characteristics											Results					
Record NO	Parent Analyte in Soil	Analyte in Soil Invertebrate	Trophic level	Tissue Type	Class	Order	Family	Genus	Species	Depurated?	Soil Conc	Soil Qualifier	Soil Units	Nominal / Measured	Tissue Conc	Tissue Unit
1	2,4-DANT	2,4-DANT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	32.4	Det	mg/kg-dw	M	165	mg/kg-dw
2	2,4-DANT	2,4-DANT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	2.17	Det	mg/kg-dw	M	5.02	mg/kg-dw
3	2,6-DANT	2,6-DANT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	44.5	Det	mg/kg-dw	M	140	mg/kg-dw
4	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Lumbricus	<i>Lumbricus terrestris</i>	No	12	Det	mg/kg-dw	M	0.140	mg/kg-dw
5	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Lumbricus	<i>Lumbricus terrestris</i>	No	61	Det	mg/kg-dw	M	2.25	mg/kg-dw
6	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	45.5	Det	mg/kg-dw	M	197	mg/kg-dw
7	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	64.9	Det	mg/kg-dw	M	296	mg/kg-dw
8	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	87.1	Det	mg/kg-dw	M	394	mg/kg-dw
9	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	109	Det	mg/kg-dw	M	591	mg/kg-dw
10	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	4.14	Det	mg/kg-dw	M	19.7	mg/kg-dw
11	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	7.69	Det	mg/kg-dw	M	39.4	mg/kg-dw
12	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	0.387	Det	mg/kg-dw	M	1.44	mg/kg-dw
13	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	1.31	Det	mg/kg-dw	M	4.39	mg/kg-dw
14	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	2.26	Det	mg/kg-dw	M	8.88	mg/kg-dw
15	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	4.68	Det	mg/kg-dw	M	19.9	mg/kg-dw
16	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	15.7	Det	mg/kg-dw	M	122	mg/kg-dw
17	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	16.1	Det	mg/kg-dw	M	157	mg/kg-dw
18	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	13	Det	mg/kg-dw	M	66.6	mg/kg-dw
19	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	15.2	Det	mg/kg-dw	M	104	mg/kg-dw
20	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	14.4	Det	mg/kg-dw	M	45.6	mg/kg-dw
21	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	50	Det	mg/kg-dw	M	37	mg/kg-dw
22	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	13	Det	mg/kg-dw	M	50.7	mg/kg-dw
23	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	11.1	Det	mg/kg-dw	M	21.6	mg/kg-dw
24	2-ADNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	4.60	Det	mg/kg-dw	M	136	mg/kg-dw
25	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Lumbricus	<i>Lumbricus terrestris</i>	No	12	Det	mg/kg-dw	M	0.150	mg/kg-dw
26	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Lumbricus	<i>Lumbricus terrestris</i>	No	91	Det	mg/kg-dw	M	1.95	mg/kg-dw
27	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	56.2	Det	mg/kg-dw	M	355	mg/kg-dw
28	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	70.4	Det	mg/kg-dw	M	414	mg/kg-dw
29	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	11	Det	mg/kg-dw	M	69	mg/kg-dw
30	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	15.6	Det	mg/kg-dw	M	96.6	mg/kg-dw
31	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	0.731	Det	mg/kg-dw	M	2.52	mg/kg-dw
32	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	2.09	Det	mg/kg-dw	M	6.58	mg/kg-dw
33	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	4.14	Det	mg/kg-dw	M	13.3	mg/kg-dw
34	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	8.75	Det	mg/kg-dw	M	30.5	mg/kg-dw
35	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	13.2	Det	mg/kg-dw	M	86.5	mg/kg-dw
36	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	14.7	Det	mg/kg-dw	M	113	mg/kg-dw
37	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	18.9	Det	mg/kg-dw	M	87.6	mg/kg-dw
38	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	21	Det	mg/kg-dw	M	104	mg/kg-dw
39	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	22.5	Det	mg/kg-dw	M	60.4	mg/kg-dw
40	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	15.3	Det	mg/kg-dw	M	121	mg/kg-dw
41	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Aporrectodea	<i>Aporrectodea rosea</i>	Yes	53	Det	mg/kg-dw	M	216	mg/kg-dw
42	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Aporrectodea	<i>Aporrectodea rosea</i>	Yes	18.2	Det	mg/kg-dw	M	25.3	mg/kg-dw
43	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	86	Det	mg/kg-dw	M	71.3	mg/kg-dw
44	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	22.8	Det	mg/kg-dw	M	51	mg/kg-dw
45	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	16.4	Det	mg/kg-dw	M	22.3	mg/kg-dw
46	4-ADNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	12.7	Det	mg/kg-dw	M	116	mg/kg-dw
47	HMX	HMX	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	9.00	Det	mg/kg-dw	M	9.00	mg/kg-dw
48	HMX	HMX	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	83	Det	mg/kg-dw	M	26	mg/kg-dw
49	HMX	HMX	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	400	Det	mg/kg-dw	N	37	mg/kg-dw
50	Perchlorate anion	Perchlorate anion	Detrivore	Whole-body	--	--	--	terrestrial	<i>terrestrial insect (misc.)</i>	--	591	Det	mg/kg-dw	M	11.6	mg/kg-dw
51	Perchlorate anion	Perchlorate anion	Detrivore	Whole-body	--	--	--	terrestrial	<i>terrestrial insect (misc.)</i>	--	42.4	Det	mg/kg-dw	M	12.4	mg/kg-dw
52	Perchlorate anion	Perchlorate anion	Detrivore	Whole-body	--	--	--	terrestrial	<i>terrestrial insect (misc.)</i>	--	42.4	Det	mg/kg-dw	M	11.4	mg/kg-dw
53	Perchlorate anion	Perchlorate anion	Detrivore	Whole-body	--	--	--	terrestrial	<i>terrestrial insect (misc.)</i>	--	3.82	Det	mg/kg-dw	M	32.3	mg/kg-dw
54	RDX	RDX	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	11	Det	mg/kg-dw	M	125	mg/kg-dw
55	RDX	RDX	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	99	Det	mg/kg-dw	M	283	mg/kg-dw
56	RDX	RDX	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	400	Det	mg/kg-dw	N	270	mg/kg-dw
57	RDX	RDX	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	100	Det	mg/kg-dw	N	240	mg/kg-dw
58	TNT	2,4-DANT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	1.36	Det	mg/kg-dw	M	5.02	mg/kg-dw
59	TNT	2,4-DANT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	2.73	Det	mg/kg-dw	M	0.836	mg/kg-dw
60	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	1.36	Det	mg/kg-dw	M	19.7	mg/kg-dw

Appendix A-2. Soil Invertebrate Bioaccumulation (Uptake) Database

Record NO	Study Location	Sub-Location	Lab / Field	Soil Characteristics														Reference
				Duration	Soil Texture	pH	OM%	CEC	Conductivity	Sulfate	%Clay	%Gravel	%Sand	%Silt	%TOC	Soil Ca		
1	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
2	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
3	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
4	--	--	Lab	>10	d	--	--	--	--	--	--	--	--	--	--	--	--	Johson et al. 1999
5	--	--	Lab	>10	d	--	--	--	--	--	--	--	--	--	--	--	--	Johson et al. 1999
6	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
7	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
8	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
9	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
10	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
11	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
12	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
13	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
14	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
15	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
16	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
17	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
18	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
19	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
20	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
21	Quebec, C B5		Field	10	d	--	5.95	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
22	Quebec, C C1		Field	10	d	--	6.81	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
23	Quebec, C C2		Field	10	d	--	6.97	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
24	Quebec, C C3		Field	10	d	--	7.03	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
25	--	--	Lab	>10	d	--	--	--	--	--	--	--	--	--	--	--	--	Johson et al. 1999
26	--	--	Lab	>10	d	--	--	--	--	--	--	--	--	--	--	--	--	Johson et al. 1999
27	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
28	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
29	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
30	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
31	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
32	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
33	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
34	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
35	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
36	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
37	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
38	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
39	Lab	--	lab	14	d	--	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
40	Quebec, C A3		Field	10	d	--	6.81	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
41	Quebec, C B2		Field	10	d	--	6.57	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
42	Quebec, C B4		Field	10	d	--	6.4	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
43	Quebec, C B5		Field	10	d	--	5.95	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
44	Quebec, C C1		Field	10	d	--	6.81	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
45	Quebec, C C2		Field	10	d	--	6.97	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
46	Quebec, C C3		Field	10	d	--	7.03	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
47	--	--	Lab	14	d	Sandy loar	5	1.3	4.27	--	--	11	--	71	18	--	--	Lachance et al. 2003
48	--	--	Lab	14	d	Sandy loar	5	1.3	4.27	--	--	11	--	71	18	--	--	Lachance et al. 2003
49	--	--	Lab	14	d	Sandy loar	5	1.3	4.27	--	--	11	--	71	18	--	--	Lachance et al. 2003
50	Mead Area		1 Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	Parsons, 2001
51	Mead Area		3 Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	Parsons, 2001
52	Mead Area		3 Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	Parsons, 2001
53	ABL		1 Field	resident	--	--	--	--	--	--	--	--	--	--	--	--	--	Parsons, 2001
54	--	--	Lab	14	d	Sandy loar	5	1.3	4.27	--	--	11	--	71	18	--	--	Lachance et al. 2003
55	--	--	Lab	14	d	Sandy loar	5	1.3	4.27	--	--	11	--	71	18	--	--	Lachance et al. 2003
56	--	--	Lab	14	d	Sandy loar	5	1.3	4.27	--	--	11	--	71	18	--	--	Lachance et al. 2003
57	--	--	Lab	14	d	Sandy loar	5	1.3	4.27	--	--	11	--	71	18	--	--	Lachance et al. 2003
58	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
59	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
60	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004

Appendix A-2. Soil Invertebrate Bioaccumulation (Uptake) Database

Record NO	Parent Analyte in Soil	Analyte in Soil Invertebrate	Soil Invertebrate Species Characteristics								Results					
			Trophic level	Tissue Type	Class	Order	Family	Genus	Species	Depurated?	Soil Conc	Soil Qualifier	Soil Units	Nominal / Measured	Tissue Conc	Tissue Unit
61	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	2.73	Det	mg/kg-dw	M	39.4	mg/kg-dw
62	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	4.50	Det	mg/kg-dw	M	1.44	mg/kg-dw
63	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	10	Det	mg/kg-dw	M	4.39	mg/kg-dw
64	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	23	Det	mg/kg-dw	M	8.88	mg/kg-dw
65	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	45	Det	mg/kg-dw	M	19.9	mg/kg-dw
66	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	99	Det	mg/kg-dw	M	122	mg/kg-dw
67	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	110	Det	mg/kg-dw	M	157	mg/kg-dw
68	TNT	2-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	135	Det	mg/kg-dw	M	66.6	mg/kg-dw
69	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	1.36	Det	mg/kg-dw	M	69	mg/kg-dw
70	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes (24 hr)	2.73	Det	mg/kg-dw	M	96.6	mg/kg-dw
71	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	4.50	Det	mg/kg-dw	M	2.52	mg/kg-dw
72	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	10	Det	mg/kg-dw	M	6.58	mg/kg-dw
73	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	23	Det	mg/kg-dw	M	13.3	mg/kg-dw
74	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	45	Det	mg/kg-dw	M	30.5	mg/kg-dw
75	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	99	Det	mg/kg-dw	M	86.5	mg/kg-dw
76	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	110	Det	mg/kg-dw	M	113	mg/kg-dw
77	TNT	4-ADNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia fetida</i>	Yes	135	Det	mg/kg-dw	M	87.6	mg/kg-dw
78	TNT	TNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Lumbricus	<i>Lumbricus terrestris</i>	No	60	Det	mg/kg-dw	M	0.0200	mg/kg-dw
79	TNT	TNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Lumbricus	<i>Lumbricus terrestris</i>	No	214	Det	mg/kg-dw	M	0.400	mg/kg-dw
80	TNT	TNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	367	Det	mg/kg-dw	M	11.9	mg/kg-dw
81	TNT	TNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	116	Det	mg/kg-dw	M	19.7	mg/kg-dw
82	TNT	TNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	130	Det	mg/kg-dw	M	10.9	mg/kg-dw
83	TNT	TNT	Detrivore	Whole-body	Oligochaeta	Haplotaxida	Lumbricidae	Eisenia	<i>Eisenia andrei</i>	Yes	116	Det	mg/kg-dw	M	11.7	mg/kg-dw

Appendix A-2. Soil Invertebrate Bioaccumulation (Uptake) Database

Record NO	Study Location	Sub-Location	Lab / Field	Duration	Soil Units	Soil Texture	Soil Characteristics										Reference	
							pH	OM%	CEC	Conductivity	Sulfate	%Clay	%Gravel	%Sand	%Silt	%TOC		Soil Ca
61	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
62	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
63	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
64	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
65	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
66	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
67	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
68	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
69	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
70	Lab	--	Lab	14	d	--	7.8	3.8	--	--	--	8	--	83	9	--	--	Lachance et al. 2004
71	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
72	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
73	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
74	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
75	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
76	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
77	--	--	Lab	14	d	Sandy	5.8	4.2	2.6	--	--	3	--	92	5	--	--	Renoux et al. 2000
78	--	--	Lab	>10	d	--	--	--	--	--	--	--	--	--	--	--	--	Johson et al. 1999
79	--	--	Lab	>10	d	--	--	--	--	--	--	--	--	--	--	--	--	Johson et al. 1999
80	Quebec, C C1		Field	10	d	--	6.81	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
81	Quebec, C C2		Field	10	d	--	6.97	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
82	Quebec, C C3		Field	10	d	--	7.03	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004
83	Quebec, C A3		Field	10	d	--	6.81	--	--	--	--	--	--	--	--	--	--	Robidoux et al. 2004

Appendix B-1
Summary of Studies Included in the Plant Bioaccumulation (Uptake)
Database

Reference:	Cataldo et al. 1993 (PNL SA-22362)
Analytes Considered:	TNT, RDX, Tetryl
Species:	Wheat and bland brome
Geographic Location of Study:	Lab
Exposure Duration:	60-d
Type of Tissue Analyzed:	Soil and various compartments in plants (leaf, stem, pod, seed, and root)
Type of Source Media Analyzed:	Soil and various plant parts
Analytical Method:	HPLC/UV
Soil Extraction Method:	Acetonitrile
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	Percent organic matter only.
Purpose of Study:	To analyze the fate of TNT, RDX and tetryl in soils and plants.
Notes:	TNT and tetryl were both metabolized a great deal in soil before they were taken up by plants. On the other hand, RDX was relatively persistent in soil. Authors found uptake of TNT was inversely related to soil percent organic matter. Since concentrations were given in wet-weight, a percent water content is assumed for each plant compartment and converted to dry-weight basis.
Reference:	Folsom et al. 1988
Analytes Considered:	TNT
Species:	Yellow nutsedge
Geographic Location of Study:	Lab
Exposure Duration:	45-d (plus 20-d pre-exposure to soil)
Type of Tissue Analyzed:	Plant leaves
Type of Source Media Analyzed:	Measured Soil and Plant leaves
Analytical Method:	Gas LC
Soil Extraction Method:	Benzene
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	Yes, available in Appendix A-1
Purpose of Study:	(1) To assess methods for soil treatment, extraction, and analyses; (2) To determine the effect of pH on TNT uptake by plants.
Notes:	Assumed soil and plant concentrations in dry-weight conc.
Reference:	Groom et al. 2002
Analytes Considered:	HMX
Species:	Native and agricultural plants
Geographic Location of Study:	Lab

Exposure Duration:	77-d
Type of Tissue Analyzed:	Aerial (leaf/stem) and root tissues
Type of Source Media Analyzed:	Soil and plants
Analytical Method:	HPLC - GC/MS
Soil Extraction Method:	Acetonitrile
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	Yes, available in Appendix A-1
Purpose of Study:	To investigate the potential of agricultural and native plants for use in phytoremediation.
Notes:	Used data from the author sent to Brad Sample, rather than estimating numbers from the figures.
Reference:	Lachance et al. 2003
Analytes Considered:	RDX, HMX, TNB, 2,4-DNT, 2,6-DNT
Species:	Alfalfa, ryegrass, Japanese millet
Geographic Location of Study:	Lab
Exposure Duration:	Various (14-d, 16-d, 19-d, 28-d, 42-d), but not uniformly applied to plant species tested.
Type of Tissue Analyzed:	Shoots
Type of Source Media Analyzed:	Mainly soil-to-plants, but has a small dataset on earthworm uptake on RDX and HMX
Analytical Method:	HPLC/UV
Soil Extraction Method:	Acetonitrile
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	pH=5.0, Sandy loam, available in Appendix A-1
Purpose of Study:	To assess how weathering of amended soil and changes in exposure concentration have on uptake potential of compounds in plants and soil invertebrates.
Notes:	All conc were in dry weight / Authors used initial measured concentration to derive BCFs, but we used final measured concentrations as this would be more realistic to field scenarios.
Reference:	Parsons et al. 2001
Analytes Considered:	Perchlorate anion
Species:	various aquatic, terrestrial plants and animals (whole body composites)
Geographic Location of Study:	Yuma, AZ/Lake Mead National Recreation Area and the Las Vegas Wash, NV / Allegany Ballistics Lab/ WV / Holloman AFB / Naval Surface Warfare Center, MD / Longhorn Army Ammunition Plant, TX
Exposure Duration:	Resident
Type of Tissue Analyzed:	Various
Type of Source Media Analyzed:	Soil, Water, Sediment
Analytical Method:	Ion chromatography and others
Soil Extraction Method:	Alumina (basic wash)

Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	No
Purpose of Study:	To determine if perchlorate are present in various biological media
Notes:	The biotic and abiotic media may not necessarily be co-located but could be grouped by location for each site.
Reference:	Price et al. 1997
Analytes Considered:	HMX, RDX, TNT
Species:	Lettuce, tomato, corn, yellow nutsedge
Geographic Location of Study:	Lab (used soil from an Army ammunition plant in Nebraska)
Exposure Duration:	45 - 80 d
Type of Tissue Analyzed:	Fruits, kernels, stovers, shoots, and leaves
Type of Source Media Analyzed:	Soil and various plant parts
Analytical Method:	HPLC/UV
Soil Extraction Method:	NA
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	Yes, available in Appendix A-1
Purpose of Study:	(1) To measure the uptake of explosives in edible plant tissues in contaminated soil. (2) To measure the effects of varying RDX and TNT in soil on plant uptake. (3) To measure uptake differences using different soil types.
Notes:	Tissue concentrations were reported in wet-weight basis; used percent water content from Table A4 to convert to dry-weight; all soil concentrations were in dry-weight, no conversion necessary. Irrigation study not incorporated as it was not applicable.
Reference:	Schneider et al. 1995
Analytes Considered:	RDX, TNT, and metabolites
Species:	Various Plants (e.g. arrowhead, corn, goldenrod)
Geographic Location of Study:	Iowa (Iowa Army Ammunition Plant)
Exposure Duration:	Resident
Type of Tissue Analyzed:	Root and Top
Type of Source Media Analyzed:	Soil and plant roots and aboveground portion
Analytical Method:	HPLC/UV
Soil Extraction Method:	Acetonitrile
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	No
Purpose of Study:	To evaluate the uptake of RDX and TNT in field plants from exposure to contaminated soil.
Notes:	
Reference:	Zellmer et al. 1995
Analytes Considered:	TNT, RDX, TNB, 2,4-DNT, 2,6-DNT, 4-ADNT, 2-ADNT

Species:	Agricultural crops (rye, oats) and various plant species
Geographic Location of Study:	Joliet Army Ammunition Plant, Joliet, IL, and lab
Exposure Duration:	Lab study: 4 and 6 months
Type of Tissue Analyzed:	Roots, shoots, and grains (seeds)
Type of Source Media Analyzed:	Soil and various plant tissues
Analytical Method:	HPLC/UV
Soil Extraction Method:	Acetonitrile
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	Soil pH, OM, CEC, and macronutrients, see Appendix A-1
Purpose of Study:	(1) To provide supporting data of the extend of TNT uptake by plants on contaminated soil; (2) To characterize how organic matter affect TNT uptake; (3) To determine the effect of organic matter on extractable TNT in contaminated soil.
Notes:	Soil concentrations were based on air-dried weight, plant concentrations presumed to be in dry-weight basis.

Appendix B-2
Summary of Studies Included in the Earthworm Bioaccumulation (Uptake)
Database

Reference:	Johnson et al. 1999
Analytes Considered:	TNT, 2-Amino-DNT, and 4-Amino-DNT
Species:	Tiger salamander and spotted salamander
Geographic Location of Study:	Lab
Exposure Duration:	28-d
Type of Tissue Analyzed:	Tissue (decapitated)
Type of Source Media Analyzed:	Soil and earthworms
Analytical Method:	NA (other than indicating Military Unique and Special Chemistry Program)
Soil Extraction Method:	NA (other than indicating Military Unique and Special Chemistry Program)
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	No
Purpose of Study:	Bioaccumulation of oral and dermal uptake to the salamanders
Notes:	Source media were analyzed at beginning, midpoint, and at the end/Estimated conc were used but ND assumed zero concentration
Reference:	Parsons et al. 2001
Analytes Considered:	Perchlorate anion
Species:	various aquatic, terrestrial plants and animals (whole body composites)
Geographic Location of Study:	Yuma, AZ/Lake Mead National Recreation Area and the Las Vegas Wash, NV / Allegany Ballistics Lab/ WV / Holloman AFB / Naval Surface Warfare Center, MD / Longhorn Army Ammunition Plant, TX
Exposure Duration:	Resident
Type of Tissue Analyzed:	Various
Type of Source Media Analyzed:	Soil, Water, Sediment
Analytical Method:	Ion chromatography and others
Soil Extraction Method:	Alumina (basic wash)
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	No
Purpose of Study:	To determine if perchlorate are present in various biological media
Notes:	The biotic and abiotic media may not necessarily be co-located but could be grouped by location for each site.
Reference:	Lachance et al. 2003
Analytes Considered:	RDX, HMX, TNB, 2,4-DNT, 2,6-DNT
Species:	Alfalfa, ryegrass, Japanese millet
Geographic Location of Study:	Lab
Exposure Duration:	Various (14-d, 16-d, 19-d, 28-d, 42-d), but not uniformly applied to plant species tested.
Type of Tissue Analyzed:	Shoots
Type of Source Media Analyzed:	Mainly soil-to-plants, but has a small dataset on earthworm uptake on RDX and HMX
Analytical Method:	HPLC/UV
Soil Extraction Method:	Acetonitrile

Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	pH=5.0, Sandy loam, see Appendix A-2
Purpose of Study:	To assess how weathering of amended soil and changes in exposure concentration have on uptake potential of compounds in plants and soil invertebrates.
Notes:	All conc were in dry weight / Authors used initial measured concentration to derive BCFs, but we used final measured concentrations as this would be more realistic to field scenarios. Lachance indicated that all concentrations were expressed in dry-weight basis (personal communication).
Reference:	Lachance et al. 2004
Analytes Considered:	TNT, 2,4-ADNT, 4-ADNT, 2,4-DANT, 2,6-DANT
Species:	Earthworm (<i>E. andrei</i>)
Geographic Location of Study:	Lab
Exposure Duration:	14-d
Type of Tissue Analyzed:	Whole-body
Type of Source Media Analyzed:	Soil
Analytical Method:	HPLC/UV
Soil Extraction Method:	Acetonitrile
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	Sandy loam forest soil, see Appendix A-2
Purpose of Study:	(1) To assess the acute toxicity of TNT and its metabolites and (2) To assess the bioaccumulation potential of TNT and its metabolites in earthworms.
Notes:	Bioaccumulation data not used from worm that were exposed to concentration beyond LC50.
Reference:	Renoux et al. 2000
Analytes Considered:	TNT, 2-ADNT, 4-ADNT, 2,4-DANT
Species:	Earthworm (<i>E. andrei</i>)
Geographic Location of Study:	Lab
Exposure Duration:	2-d and 14-d, followed by 24 hr depuration
Type of Tissue Analyzed:	Whole-body
Type of Source Media Analyzed:	Forest soil
Analytical Method:	HPLC/UV
Soil Extraction Method:	Acetonitrile
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	Available

Purpose of Study:	(1) To assess the ability of earthworm to biodegrade TNT, and (2) to characterize the extend of TNT biodegradation by earthworms by analyzing the distribution of TNT and its metabolite in soil and in earthworm and by identifying the enzymatic activity of the earthworms.
Notes:	Bioaccumulation data not used from worm that were exposed to concentration beyond LC50.
Reference:	Robidoux et al. 2004
Analytes Considered:	TNT, 4-ADNT, and 2-ADNT
Species:	Earthworm (E. andrei and A. rosea)
Geographic Location of Study:	Lab
Exposure Duration:	10-d, followed by 8 hr depuration
Type of Tissue Analyzed:	Whold-body
Type of Source Media Analyzed:	Soil and earthworms
Analytical Method:	HPLC/UV
Soil Extraction Method:	Acetonitrile
Soil Characteristics (pH, CEC, % OM, % Clay, etc.) Presented:	pH and water holding capacity available
Purpose of Study:	To assess the toxicity of TNT in soil to earthworms, and to measure biomarks of exposure, including tissue residues in mesocosms.
Notes:	Authors found TNT metabolites in earthworms not correlated with TNT soil concentrations.